Effect of NaCl and Phenylalanine on the Production of some Secondary Metabolites in *In Vitro* Cultures of *Mentha longifolia*

Ola A. EL-Shennawy, Mohamed G. El-Torky, Hoda E. El-Mokadem and Basem I. Abass¹

ABSTRACT

The present study was conducted at the Tissue Culture and Plant Micropropagation Laboratory of Floriculture, Ornamental Horticulture and Landscape Gardening Department, Faculty of Agriculture, Alexandria University, during 2014 and 2015, to study the effect of five different concentrations of phenylalanine (0, 0.5, 5, 10, and 15 mg/l) and four different concentrations of sodium chloride (0, 2000, 4000 and 6000 mg/l) on the production of some secondary metabolites in *in vitro* cultures of *Mentha longifolia* using shoot tips and leaves.

Leaf segments about 1 cm² and shoot tips consisting of 3-4 cm long stem segments containing three auxiliary buds (as explants) were planted on MS medium supplemented with NAA (0.25 mg/l) + BAP(2.5 mg/l) + 30 g/liter sucrose + 7 g/liter agar for shoot regeneration from shoot tips, while MS medium contained 2,4-D (4 mg/l) + BAP (0.5 mg/l) for callus induction from leaf segments, the pH was adjusted to 5.8±0.02. Plantlets were subjected to 16 hours light and 8 hours darkness (2000- 2500 lux for 16 h/d provided by fluorescent tubes), the temperature was adjusted at 25° C \pm 2 and 60-70% humidity. The cultures were incubated for 60 days in case of shoot tips and 100 days in case of leaves for callus induction. Chemical analysis was performed on callus tissues after 60 and 100 days from planting and shoot tissues after 60 days from planting which were produced under salinity levels, phenylalanine levels and their interaction to determine the contents of total chlorophyll, total phenolic, rosmarinic acid and proline in both tissues.

From the obtained results in this study, using shoot tips for the production of phenolic acid and rosmarinic acid was superior than the callus. Low concentration of sodium chloride (2000 mg /l) enhanced the production of some secondary metabolites. However high concentrations of sodium chloride (6000 mg /l) leads to slow growth, yellowing of cells and sometimes the deterioration of plants. Uses of intermediate concentrations of phenylalanine (5 and 10 mg /l) enhanced the production of chlorophyll and some secondary metabolites where as high concentration (15 mg /l) does not stop growth and cell division.

Key words: *Mentha longifolia* – Amino acids – *In vitro* – Secondary metabolites – Salt stress – Lamiaceae.

INTRODUCTION

Medicinal plants are an important source of drugs that had been used thousands of years ago (Clark, 1996). With the advent of synthetic drugs and replacing herbal medicines with them, the medicinal plants gradually faded. Then chemical drugs and antibiotics in the treatment of a wide variety of diseases due to the high speed of treatment were used. Due to harmful side effects of chemical drugs for human health, in recent years, attention and requesting of people to use herbal medicines to treat diseases has increased. As a third to a half of pharmaceutical products in United States of America are plant-derived (Molsaghi *et al.*, 2014). In recent years, researches on the effects of natural inhibition against microorganisms showed that herbal products should be alternatives to synthetic drugs and may have a significant therapeutic effect (Nassiri and Hosseinzadeh, 2008).

Mentha longifolia (L) Hudson (wild mint, English horsemint) belongs to the genus Mentha, Lamiaceae family and is associated with medicinal and aromatic herbs. It is widespread throughout the Mediterranean, Central and Northern Europe, Asia Minor and Africa (Janković 1974 and Stamenković, 2005). Approximately 25 species belongs to Genus Mentha. Medicinal importance of Mentha is well known due to the presence of rosmarinic, the second most common ester of caffeic acid in the plant kingdom (Ellis and Towers, 1970). Herbal polyphenolic compounds are secondary metabolites with a characteristic aromatic structure, which can be classified into fifteen groups according to the basic part of their molecule, as for example phenols, phenolic acid, flavonoids, anthocyanins, quinones, catechins and tannins, just to name a few (Dilas et al., 2002). Rosmarinic acid (RA), exhibits various pharmacological activities including prevention of oxidation of low density lipoprotein and anti-allergic action. The biological activity of RA is described as antibacterial, antiviral, and antioxidative (Szabo et al., 1999).

Mentha longifolia is used in the pharmaceutical, tobacco, food industries and particularly in cosmetology. Different parts of the plant including its leaves, flower, stem, and seeds have been also used widely in traditional folk medicine as antimicrobial, carminative, stimulant, antispasmodic and for the treatment of various diseases such as headaches and digestive disorders (Naghibi *et al.*, 2005). In pharmacological research, there is enough indication for

¹ Department of Floriculture, Ornamental Horticulture and Landscape Gardening, Faculty of Agriculture, Alexandria University. Received July 10, 2017, Accepted September 20, 2017

different biological effects of *M. longifolia* and the chemical compounds present in the essential oil of the plant.

Plant tissue culture, or the aseptic culture of cells, tissues, organs, and their components under defined physical and chemical conditions in vitro, is an important tool in both basic and applied studies as well as in commercial application. It owes its origin to the ideas of the German scientist, Haberlandt, at the begining of the 20th century. The early studies led to root cultures, embryo cultures, and the first true callus/tissue cultures. The period between the 1940s and the 1960s was marked by the development of new techniques and the improvement of those that were already in use. It was the availability of these techniques that led to the application of tissue culture to five broad areas, namely, cell behavior (including cytology, nutrition, metabolism, morphogenesis, embryogenesis, and pathology), plant modification and improvement, pathogen free plants and germplasm storage, clonal propagation, and product (mainly secondary metabolite) formation, starting in the mid-1960s. The 1990s saw continued expansion in the application of the in vitro technologies to an increasing number of plant species. Cell cultures have remained an important tool in the study of basic areas of plant biology and biochemistry and have assumed major significance in studies in molecular biology and agricultural biotechnology (Thorpe, 2007).

Plants are usually exposed to different environmental stresses which limit their growth and productivity as well as cause considerable loss to worldwide agricultural production (Shao *et al.*, 2009). One of the most important factors affecting plant growth and the production of secondary metabolites is the salt stress (Nikolova and Ivancheva, 2005). Salinity of soil or water is one of major stress obstacles to increase production in plant growing areas throughout the world and especially in arid and semi-arid regions it can severely limit plant production (Jamil *et al.*, 2006). About 23% of the world's cultivated lands is saline and 37% is sodic (Khan and Duke, 2001).

Sodium chloride and some amino acids could promote *in vitro* production of some plant secondary metabolites. Secondary metabolites are often referred to as compounds that have no fundamental role in the maintenance of life processes in the plants, but they are important for the plant to interact with its environment for adaptation and defense.

Therefore, the objective of this study is enhancing some secondary products such as total phenolic compounds and rosmarinic acid in *Mentha longifolia* through salt stress and phenylalanine *in vitro* using leaves and shoot tips explants.

MATERIALS AND METHODS

The experiment was performed at the Tissue Culture and Plant Micropropagation Laboratory of Floriculture, Ornamental Horticulture and Landscape Gardening Department, Faculty of Agriculture, Alexandria University, during 2014 and 2015.

of Source plant material and explants sterilization: Two months old potted plants of Mentha longifolia L. were obtained from a commercial nursery. Explants consisting of 3-4 cm long stem segments containing three auxiliary buds and leaf segments (1 cm²) were used. The explants were washed with tap water, they were surface sterilized with 70% ethanol for 1 min followed by soaking in 0.1% mercuric chloride and 2-3 drops of Tween 20 for 1-2 min and finally washed thoroughly 4-5 times in a sterile double distilled water to remove any traces of mercuric chloride. Explants were cut into small pieces and cultured in 200 ml jars containing 20 ml of culture medium.

Media preparation: Murashige and Skoog (MS) basal medium with vitamins + 30 g/liter sucrose + 7 g/liter agar was used. The pH was adjusted to 5.8 ± 0.02 using 1N NaOH and 1N HCL. Medium was then poured in glass jars (200 ml) then autoclaved for 20 minutes at 121 °C and 120 bar/cm² (Mehta *et al.*, 2012).

MS medium supplemented with NAA (0.25 mg/l) and BAP (2.5 mg/l) was used for shoot tips regeneration (Roy and Mukhopadhyay, 2012). Five concentrations of phenylalanine (0, 0.5, 5, 10, and 15 mg/l) and four concentrations of NaCl (0, 2000, 4000 and 6000 mg/l) were added to the growing media. The cultures were subjected to 16 hours light and 8 hours darkness (2000-2500 lux for 16 h/d provided by fluorescent tubes), the temperature was adjusted at 25° C \pm 2 and 60-70% relative humidity. Callus was induced on MS medium containing both 2, 4-D (4 mg/l) and BAP (0.5 mg/l) (Darvishi *et al.* 2014).

Collected data: Chemical analysis was performed on both shootlets and callus tissue after 60 days and 100 days only in callus tissue, to determine:

Total chlorophyll content: Total chlorophyll was determined as (μ g/g FW) according to Moran and Porath (1980).

Total phenolic compounds: The free bound phenolic contents in shootlets and callus were determined using folin- ciocalteu's method (AOAC, 2000) as (mg/g FW).

Rosmarinic acid (RA): The method of RA extraction used through this study was reported by Lopez-Arnoldos *et al.* (1995) and Komali and Shetty (1998) as (µmol/g FW).

Proline content: Proline was estimated using the acid ninhydrin method as described by Bates *et al.* (1973) as $(\mu g/g \text{ FW})$.

Statistical analysis: The experiment was designed as factorial in a split plot design with two factors, where phenylalanine treatments (0, 0.5, 5, 10, and 15 mg/l) were arranged as main plots and sodium chloride treatments (0, 2000, 4000 and 6000 mg/l) were set as sub plots over two explants (shoot tips and leaves) each with 3 replicates. Data were analyzed for

significance by analysis of variance according to Steel and Torrie (1980). Statistical analysis was done by ANOVA, F-test, and L.S.D procedures available within the SAS software package (Version 9.13 2007).

RESULTS AND DISCUSSION

Effects of phenylalanine on chlorophylls content of *Mentha longifolia* explants

Generally, data presented in Table (1) revealed that adding phenylalanine alone to the medium caused significant increasing in chlorophylls (a, b and total) content of the used explants i.e. callus and shootlets, compared with the control treatment (medium free from phenylalanine). Also, using phenylalanine alone at 5 mg/l gave the highest significant average mean of total chlorophyll of callus and / or shootlets, compared with the other concentrations. The previous treatment led to increase the total chlorophyll content of callus with 64.09 % after 60 days and with 90.56 % after 100 days and with 51.40 % for shootlets after 60 days over the control treatment. Besides, increasing the concentration of phenylalanine over 5 mg/l in the media led to significant decrease the total chlorophyll content of the used explants.

These results were probably attributed to the direct role of the amino acid phenylalanine in the biosynthesis of the green pigments, which considers as a source for nitrogen and carbon as structural components of chlorophyll formation, hence adding phenylalanine at a proper concentration to the medium led to activate chlorophyll synthesis and protect its decomposition, consequently its content in the used explants could be increased.

Our results are similar to the findings of Ghai *et al.* (2002) on *Brassica napus*, Khodary (2004) on *Zea mays*, Agarwal *et al.* (2005) on wheat, El-Tayeb (2005) on barely, and Hayat and Ahmad (2007).

Effects of sodium chloride on chlorophylls content of *Mentha longifolia* explants

Generally, data presented in Table (2) showed that adding sodium chloride at (4000 and 6000) mg/l to the medium gave the highest significant average value of total chlorophyll content of *Mentha longifolia* callus had a high resistance for salt stress, thus increasing sodium chloride in the medium led to increase the total chlorophyll content in their tissues.

Marcelis and Hooijdonk, (1999) reported that salinity reduces the chlorophyll content in salt susceptible plants and increases it in salt tolerant plants as for callus where increased salinity levels caused an increase in chlorophyll a and b contents and total chlorophyll.

Besides, data in Table (2) revealed that using sodium chloride at any concentration in the medium led to significant decrease the average mean of total chlorophyll content of *Mentha longifolia* shootlets compared with the control treatment (medium free of sodium chloride).

Weimberg (1975), Munjal and Goswami (1995), Soussi et al. (1998), Jenifer and Franklin- Janus (2002) and Xu et al. (2008) reported that salinity caused a decrease in chlorophyll and carotenoids level in other plants. Decrease in chlorophylls level under salt stress may be due to reduction in pigment biosynthesis or enzymatic chlorophyll degradation (Xu et al., 2008 and Yang et al., 2009). The loss of chlorophyll under salt stress could be related to photoinhibition or reactive oxvgen species formation. The reduction in photosynthesis under salinity can also be attributed to a decrease in chlorophyll content. (Kato and Shimizu, 1985). The observed decrease of chlorophyll content in the plants grown under saline conditions may be attributed both to the increased degradation and the inhibited synthesis of that pigment (Garsia-Sanchez et al., 2002). Moreover, the reduction in photosynthetic pigments (chlorophyll a, chlorophyll b, and total carotenoids) due to salinity effect which attributed to the disturbance of ions absorption involved in chloroplast formation and protein synthesis and/or plastid breakdown (Abd El-Wahab, 2006). However, it was observed that the degradation of chlorophyll pigments was increased as a result of exposure to NaCl (Eryilmaz, 2006).

The present results are in agreement with those of Siler *et al.* (2007), on centaury, Koocheki *et al.* (2008), on *Teucrium polium*, *Thymus vulgaris*, *Zataria multiflora*, and *Ziziphora clinopodioides*, and Najafi *et al.* (2010), on *Satureja hortensis*.

Effect of the interaction between phenylalanine and sodium chloride on contents of *Mentha longifolia* explants of chlorophyll a, chlorophyll b and total chlorophyll

Data in Table (3) showed that medium with 5 mg/l Phenylalanine and free from sodium chloride gave the highest values of total chlorophyll content (390.79) in

Table 1. Means of c	hlorophyll a (Chl intrations	l.A), chlorophyl	l b (Chl.B) and	total chlorophy	/ll content (T.C	hl) of callus and	1 shootlets of M	lentha longifoli	a as affected by
Phenylalanine		Callus			Callus			Shootlets	
mg/l		60 days			100 days			60 days	
		Chl.B	T.Chl	Chl.A	Chl.B	T.Chl	Chl.A	Chl.B	T.Chl
	CIII.A jug/g	hg/g	b'Bn	hg/g	a/Bri	a/Bri	a/an	hg/g	b/an
0	11.276 c	8.132 c	19.408 c	10.217 b	5.346 b	15.563 c	145.34 c	37.610 b	182.95 bc
0.5	14.628 b	11.162 ab	25.790 b	16.043 a	10.360 a	26.403 ab	189.77 b	51.694 a	241.464 a
S	19.594 a	12.253 a	31.847 a	17.453 a	12.204 a	29.657 a	224.06 a	52.932 a	276.992 a
10	4.297 e	12.033 a	16.330 d	10.444 b	6.117 b	16.561 c	146.65 c	40.681 b	187.331 b
15	8.198 d	9.496 bc	17.694 cd	11.602 b	10.425 a	22.027 b	111.87 d	37.479 b	149.349 c
L.S.D (0.05)	1.685	1.701	3.0199	2.390	2.838	4.387	28.956	8.2172	36.586
L.S.D: least significant of Means followed by the s	differences at 0.05 of same letter(s) in each	probability. column are not si	gnificantly different	t according to L.S.	D at 0.05 level of	probability.			
Table 2. Means of ch as affected by sodiur	nlorophyll a (Chl. n chloride (NaCl)	A), chlorophyll concentrations	b (Chl.B) and to	tal chlorophyll (content (T.Chl)	(µg/g fresh weig	ht) of callus and	d shootlets of M	fentha longifolia
NaCl		Callus			Callus			Shootlets	
mg/l		60 days			100 days			60 days	
	Chl.A	Chl.B	T.Chl	Chl.A	Chl.B	T.Chl	Chl.A	Chl.B	T.Chl
	h8/8	lug/g	g/gu	ng/g	lug/g	ug/g	hã/ã	ug/g	ng/g
0	2 CEL 8	7 714 h	16 446 d	11 340 0	7 775 h	126150	250 82 9	75 577 9	335 40 9

	σ	-
2	he	al
	ny	le
	al	÷
	an	M
•	ine	ear
	CO	IS (
	nc	OI C
	ent	chi
	rat	oro
	ior	ph
	S	УII
		2
		Ch
		1.2
		9,
2		chi
		orc
		ndc
		IVI
		0
		0
		hl.1
		9
		and
		d tu
		ota
		- Cl
		10
		lou
		yuc
		111
		noo
2		ter
		11
		T.C
		IU
) 0
		T C2
		allu
		IS SI
		ma
		SL
		100
		TIE
		C SI
		U IC
		Viet
2		un
		alu
-		Suo
-		ofic
		la
		as
		at
		Tect
		ted
		0

a

and for managerin on	THE CHICK INC. I. THE	ci) concenti ation	5						
NaCl		Callus			Callus			Shootlets	
mg/l		60 days			100 days			60 days	
	Chl.A	Chl.B	T.Chl	Chl.A	Chl.B	T.Chl	Chl.A	Chl.B	T.Chl
ĺ	ng/g	ng/g	ng/g	ng/g	ng/g	ha/a	ng/g	ng/g	ng/g
0	8.732 c	7.714 b	16.446 d	11.340 c	7.275 b	18.615 c	259.82 a	75.577 a	335.40 a
2000	10.103 c	8.783 b	18.886 c	14.223 b	7.171 b	21.394 b	145.33 b	38.115 b	183.42 b
4000	15.688 a	13.417 a	29.105 a	9.674 c	7.566 b	17.240 c	152.81 b	35.682 b	188.49 b
6000	11.870 b	12.547 a	24.419 b	17.369 a	13.547 a	30.916 a	96.20 c	26.944 c	123.14 c
L.S.D (0.05)	1.422	1.312	2.321	1.845	1.474	2.468	27.017	8.138	34.051
	1.00								

L.S.D: least significant differences at 0.05 of probability. Means followed by the same letter(s) in each column are not significantly different according to L.S.D at 0.05 level of probability.

L.S.D: least			Shootlets					Callus						Explant typ	Table 3. N sodium ch
(0.05) significar	15	10	S	0.5	0	15	10	S	0.5	0	mg/l	Phe		ē	leans of loride (N
t differenc	267	327.52	390.79	381.61	310.06	11.34	10.83	25.75	19.69	14.59	0		(Tota	total chlo IaCl) con
52	132	147.56	300.30	195.24	142.11	13.81	12.40	27.21	23.53	17.69	2000	NaC	ıg/g fresh v	chlorophy	rophyll, p centration
.847	83.09	154.32	248.69	280.59	175.73	45.61	16.67	33.46	24.14	25.63	4000	l mg/l	veight)	Il content	henols, re 18
	115.28	119.91	168.19	108.33	103.90	0000	25.40	41.18	35.78	19.70	6000				osmarinic
	6.37	6.35	6.03	4.60	4.81	5.92	4.76	2.40	1.72	5.43	•				acid and
1.65	6.61	6.17	6.49	2.89	7.69	5.58	4.87	1.26	2.38	6.22	2000	NaCI	(mg/g fres	Total pl	proline co
48	5.22	5.48	3.72	5.66	3.06	5.56	5.26	2.98	1.63	6.30	4000	mg/l	1 weight)	nenols	ntents of
	4.83	4.73	4.61	3.23	3.97	0000	5.55	4.42	4.47	4.79	6000				f callus :
	1.484	1.354	1.168	0.848	0.669	1.074	0.750	1.026	0.435	0.730	0		(j	Ro	and sho
0.4	1.477	1.282	1.131	0.371	1.908	1.060	0.893	0.097	0.288	0.812	2000	NaCI	umol/g fro	smarinic	tlets of .
802	0.757	1.009	0.595	0.970	0.468	0.967	0.935	1.390	0.192	0.916	4000	mg/l	esh weigl	acid cont	Mentha
	0.834	0.731	0.642	0.515	0.583	0000	1.146	1.028	1.095	0.757	6000		lt)	ent	longifoli
	75.45	111.57	87.06	72.23	70.29	102.85	338.85	281.54	61.29	1303.82	0				a as affec
194.	67.07	142.53	75.45	96.09	129.63	211.93	713.72	109.08	46.74	545.42	2000	NaCi	(µg/g fre	Proline	ted by ph
6157	74.81	162.52	64.49	70.29	83.19	1386.93	243.10	172.45	25.966	68.56	4000	l mg∕l	sh weight)	• content	enylalanii
	80.61	98.03	34.82	107.70	87.06	261.80	1106.43	398.93	430.10	71.68	6000				ıe (Phe) ar

SOC	Ta
Ī	ble
BC	\$
hlo	Me
Ē	ans
e (i	of
a	to
3	al
<u>10</u>	chl
ICe	oro
ħ	ph
ati	y∥,
ons	ph
-	iene
	ols,
	ro
	sma
	arii
	nic
	aci
	da
	nd
	pro
	Dlin
	le c
	ont
	ent
	S O
	fca
	IIu
	s a
	nd
	sho
	otl
	ets
	of
	Me
	nth
	ale
	Suc
	ifo
	lia
	as
	Iffe
	cte
	d b
	Уp
	hei
	ilýt
	alaı
	nin
	e (I
	he
) aı
	nd

shootlets, while medium with 10 mg/l Phenylalanine and free from sodium chloride gave the lowest value of total chlorophyll content (10.83) in callus.

Generally, data presented in Table (3) showed that adding phenylalanine at 0.5 and / or 5 mg/l to the medium in absence of sodium chloride gave the maximum significant content of total chlorophyll in the shootlets of *Mentha longifolia*, compared with the other explant (callus) and / or concentrations.

These results may be due to that the shootlets of *Mentha longifolia* are considered as salt sensitive and adding phenylalanine at a proper concentration to the medium led to activate the chlorophyll synthesis in their tissues.

Also, data in Table (3) indicated that adding sodium chloride at 2000 mg/l to the medium in absence of phenylalanine gave the highest significant values of total phenols and rosmarinic acid in the shootlets of *Mentha longifolia*, compared with the other explant (callus) and / or concentrations.

These results may be due to that the exposure of *Mentha longifolia* shootlets to salinity stress at a specific concentration led to force their tissues to form the antioxidant materials which are important for plant protection.

In addition, results in Table (3) revealed that adding sodium chloride at 4000 mg/l combined with phenylalanine at 15 mg/l to the medium gave the highest significant value of proline in the callus tissues of *Mentha longifolia*, compared with the other explant and / or concentrations.

In the present study generally, sodium chloride caused an increase in proline content of Mentha longifolia shootlets and callus in comparison to the control. Proline is an amino acid that increases its value in tissue of plants, under environmental stress conditions (Aziz et al., 2008). The increase in proline content could be attributed to a decrease in proline oxidase activity in saline conditions (Muthukumarasamy et al., 2000). Plants under salt stress may present increased levels of certain compounds. Amino acids (alanine, arginine, glycine, serine, leucine, and valine, together with the amino acid, proline, and the non-protein amino acids, citrulline and ornithine) and amides (such as glutamine and asparagines) have also been reported to accumulate in plants subjected to salt stress (Mansour, 2000). On Catharanthus roseus and Matricaria chamomilla higher total free amino acids in plants subjected to salt stress conditions were reported by Osman et al. (2007) and Cik et al. (2009).

Proline may be act as radical scavenger and protects cells against salt induced oxidative stress (Hong et al., 2000). The increase of free proline under salt stress may be due to the stimulation of its synthesis which is a useful defensive way of plants (Saliem, 2000). Also the increase of free proline is a result of osmotic disturbance within cells that leads to a decrease of the osmotic potential and proline is synthesized in the cytoplasm of stressed cells which keep the equilibrium between vacuole and cytoplasm (Delauney and Verma, 1993). Further, the accumulation of free proline under high sodium chloride and high exogenous proline may be related to the synergistic effect of both of them. Proline plays an essential role in the osmoregulation of plant cells when it accumulates at high concentration at the cvtoplasm and decreases water potential of cytoplasm and this will cause a balance with the low water potential of the vacuole resulting from the accumulation of ions in it and that will keep a suitable turgor of cells and grow under salt stress condition (Greenway and Munns, 1980).

Our results are in agreement with those of Abraham *et al.* (2003) who reported that proline occurs widely in higher plants and accumulates in larger amounts than other amino acids. According to literature, proline accumulates in leaves as a response to salt stress by Hendawy and Khalid (2005) on *Salvia officinalis*, Ashraf and Orooj (2006) on *Trachyspermum ammi*, Al-Amier and Craker (2007) on spearmint, Ali *et al.* (2007) on chamomile and sweet marjoram, Osman *et al.* (2007) on *Catharanthus roseus*, Abd EL-Azim and Ahmed (2009) on *Achillea fragratissima*, Cik *et al.* (2009) on *Matricaria chamomilla*, Zaki *et al.* (2009) on sweet fennel and Najafi *et al.* (2010) on *Satureja hortensis*.

Effects of phenylalanine on total phenolic compounds and rosmarinic acid content of *Mentha longifolia* explants

Generally, data presented in Table (4) on the averages values of the total phenolic compounds and rosmarinic acid content of the periods (60 and 100 days) indicated that adding phenylalanine at 5 mg/l to the medium gave the highest values of the two compounds in *mentha longifolia* callus, compared with the other concentrations. While as for the shootlets, data in Table (4) showed that using phenylalanine at 15 mg/l gave the highest content of total phenols and rosmarinic acid in their tissues compared with the other concentrations.

These results were probably due to that each plant tissue requires a proper concentration from phenylalanine to stimulate of key enzyme Phenylalanine Ammonia Lyase (PAL) activity which is the gateway of shikimik acid pathway,

shootlets of	Mentha longifolia	as affected by ph	renylalanine conc	entrations					
phenols		Cal	lus		Sho	otlets	0	allus	Shootlets
mg/l	b 09	ays	100 d	ays	60	days	60 days	100 days	60 days
	T.Phenols	RA	T.Phenols	RA	T.Phenols	RA	Prolin	e content	Proline content
	mg/g	jumol/g	mg/g	jumo]/g	mg/g	jumol/g	_	(g/gu	(jug/g)
0	5.687 a	0.804 a	3.144 c	0.476 d	4.889 c	0.907 b	497.37 a	424.65 a	92.549 b
0.5	2.554 c	0.503 b	5.709 a	0.884 b	4.100 d	0.676 c	141.03 b	224.66 b	86.583 b
S	2.770 c	0.885 a	6.277 a	1.080 a	5.219 bc	0.884 b	240.50 b	123.10 b	65.461 c
10	5.116 a	0.931 a	2.903 c	0.597 cd	5.688 ab	1.092 a	600.53 a	221.28 b	128.666 a
15	4.269 b	0.775 a	4.659 b	0.778 bc	5.760 a	1.138 a	490.88 a	203.36 b	74.491 c
L.S.D (0.05)	0.781	0.247	0.834	0.182	0.4831	0.1315	169.49	132.77	10.158
Table 5. Me shootlets of	ans of total phen Mentha longifolia	olic compounds (t as affected by so	T.Phenols) (mg/g dium chloride (N	fresh weight) a aCl) concentrat	nd contents of r ions	osmarinic acid	l (RA) (µmol/g fre	esh weight) and J	proline of callus and
NaCl		C	allus		Shooth	ets	Callu		Shootlets
mg/l		60 days	100	days	60 day	15	60 days	100 days	60 days
	T.Pheno	ls RA	T.Phenols	RA	T.Phenols	RA	Proline co	ntent	Proline content
	mg/g	jumol/g	mg/g	jumol/g	mg/g	µmol/g	a/an		g/Bri
0	4.052 a	0.803 ab	3.988 b	0.654 bc	5.637 a	1.105 a	417.67 ab	160.19 b	83.326 c
2000	4.065 a	0.630 b	3.738 b	0.564 c	5.975 a	1.234 a	325.38 b	157.08 b	102.159 a
4000	4.351 a	0.880 a	5.016 a	0.740 b	4.635 b	0.759 b	379.40 ab	360.49 a	91.065 b
6000	3.849 a	0.805 ab	5.412 a	1.094 a	4.279 b	0 661 h	453 70 a	070 88 9	81.649 c
H D						01001.0	TJJ.17 G	117.00 G	

 L.S.D (0.05)
 NS
 0.223
 0.213

 L.S.D: least significant differences at 0.05 of probability.
 0.800
 0.900

 N S: Not significant at 0.05 level of probability.
 NS: Not significant at 0.05 level of probability.

 Means followed by the same letter(s) in each column are not significantly different according to L.S.D at 0.05 level of probability.

which in turn produced the phenolic compounds as reported by Razzaque and Ellis (1977).

Similar trend of results was reported by Phatak and Heble (2002), Kim *et al.* (2006), Namdeo *et al.* (2007) and Roy and Mukhopadhyay (2012).

Furthermore, data on proline content in Table (4) revealed that tissue callus contained more proline than that of shootlets, and adding phenylalanine alone at any concentration to the growing medium led to significant decrease the proline content of the used explants, compared with medium free from phenylalanine, with one exception of using phenylalanine at 10 mg/l.

These results may be due to that adding phenylalanine at a proper concentration to the medium led to activate the growth of the used explants and reduce their stress, consequently their proline content could be decreased.

Effects of sodium chloride on total phenolic compounds and rosmarinic acid content of *Mentha longifolia* explants

Generally, data presented in Table (5) indicated that adding sodium chloride to the growing medium at 4000 or 6000 mg/l for callus or at 2000 mg/l for the shootlets gave the highest average contents of total phenolic and rosmarinic acid, compared with the control treatment.

These results were probably due to that adding sodium chloride at a specific concentration for each plant tissue led to exposure the used explant to salt stress, consequently the explant tried to avoid harmful effect of salinity by increasing the formation of some organic materials such as phenols and rosmarinic acid.

Nikolova and Ivancheva (2005) reported that the content of some secondary plant products are significantly higher in plants grown under salt stress than in those cultivated under normal conditions. The present results are in agreement with those of Al-Amier and Craker (2007) on spearmint they found that phenolic acid concentration increased in salt stressed plants and Abd EL-Azim and Ahmed (2009) they indicated that the content of phenols on Achillea fragratissima significantly increased along with increasing salinity. Also, Cik et al. (2009) found that accumulation of phenolic acids (protocatechuic, chlorogenic and caffeic acids) on Matricaria chamomilla increased by salinity. Bourgou et al. (2010) showed increase of phenols with saline treatment on Nigella sativa and Queslati et al. (2010) on Mentha pulegium.

Effects of sodium chloride on proline content of *Mentha longifolia* explants

Data in Table (5) showed that increasing the added sodium chloride to the growing medium led to increase the proline content in each of callus and / or shootlets tissues of *Mentha longifolia*, compared with the control treatment (medium free from sodium chloride).

These results may be attributed to the importance of presence of proline at a proper level in plant tissues during salt stress, which in turn imparts stress tolerance by maintaining cell turgor, or osmosic balance, stabilizing memberanes thereby preventing electrolyte of reactive oxygen species within normal ranges, thus preventing oxidative burst in plants as reported by Hayat *et al.* (2012).

REFERENCES

- Abd EL-Azim, W. M. and S. Th. Ahmed. 2009. Effect of salinity and cutting date on growth and chemical constituents of *Achillea fragratissima* Forssk, under Ras Sudr conditions. Res. J. Agric. Biol. Sci., 5(6): 1121-9.
- Abd El-Wahab, M. A. 2006. The efficiency of using saline and fresh water irrigation as alternating methods of irrigation on the productivity of *Foeniculum vulgare* Mill subsp. vulgare var. vulgare under North Sinai conditions. Res. J. Agr. Biol. Sci., 2(6): 571-7.
- Abraham, E., G. Rigo, G. Szekely, R. Nagy, C. Koncz and L. Szabados. 2003. Light-dependent induction of proline biosynthesis by abscisic acid and salt stress is inhibited by brassinosteroid in Arabidopsis. Plant Mol. Biol., 51: 363-72.
- Agarwal, S., R. K. Sairam, G. C. Srivatava, A. Tyagi and R. C. Meena. 2005. Role of ABA, salicylic acid, calcium and hydrogen peroxide on antioxidant enzymes induction in wheat seedlings. Plant Science., 169: 559–570.
- Al-Amier, H. and L. E. Craker. 2007. *In vitro* selection for stress tolerant spearmint. reprinted from: Issues in New Crops and New Uses. J. Janick and A. Whipkey (eds.). ASHS Press, Alexandria, VA. 306-10.
- Ali, R. M., H. M. Abbas and R. K. Kamal. 2007. The effects of treatment with polyamines on dry matter, oil and flavonoid contents in salinity stressed chamomile and sweet marjoram. Plant Soil Envir., 53: 529-43.
- AOAC, 2000. Official Methods of Analysis (17th ed) Association of Official Analytical Chemists. Boon, H. and M. Smith, 1999. The Botanical Pharmacy. The Pharmacy. The Pharmacology of 47 Common Herbs, 249-296.
- Ashraf, M. and A. Orooj. 2006. Salt stress effects on growth, ion accumulation and seed oil concentration in an arid zone traditional medicinal plant ajwain (*Trachyspermum ammi* [L.] Sprague). J. Arid. Env., 64(2): 209-20.
- Aziz, E. A., H. Al-Amier and L. E. Craker. 2008. Influence of salt stress on growth and essential oil production in Peppermint, Pennyroyal and Apple Mint. J. Herbs, Spices and Med. Plants., 14: 77-87.

- Bates, L. S., Waldren, R. P. and Teare, I. D. 1973. Rapid determination of free proline for water-stress studies. Plant and Soil. 39(1): 205-207.
- Bidwell, R. G. S. 1979. Plant Physiology, 2nd ed. MacMillan Publish., New York.
- Bourgou, S., M. E. Kchouk, A. Bellila and B. Marzouk. 2010. Effect of salinity on phenolic composition and biological activity of *Nigella sativa*. Acta Hort. 853: 57-60.
- Cik, J. K., B. Klejdus, J. Hedbavny and M. Bačkor. 2009. Salicylic acid alleviates NaCl-induced changes in the metabolism of *Matricaria chamomilla* plants. Ecotoxicology. 18(5): 544-54.
- Clark, A. M. 1996. Natural products as a resource for new drugs. Pharm. Res.13: 1133-1141.
- Darvishi, E., E. Elham Kazemi, D. Kahrizi, S., Bahraminejad, M. Mansouri, S. R. Chaghakaboudi and Y. Khani 2014. Optimization of callus induction in pennyroyal (*Mentha pulegium*). J. Appl. Biotech. Reports., 1(3): 97-100.
- Delauney, A. and D. Verma. 1993. Proline biosynthesis and osmoregulation in plants. The Plant J., 4(2): 215-223.
- Đilas, S. M., J. M. Čanadanović-Brunet and G. R. Četković. 2002. Antioxidants in food, Chem. Ind., 56: (3) 105-112.
- Ellis, B. E. and G. H. N. Towers. 1970. Biogenesis of rosmarinic acid in *Mentha*. Biochem. J., 118: 291-297.
- El-Tayeb, M. A. 2005. Response of barley grain to the interactive effect of salinity and salicylic acid. Plant Growth Regulation., 42: 215–224.
- Eryilmaz, F. 2006. The relationships between salt stress and anthocyanin content in higher plants. Biotech. and Biotech. Eq., 20(1): 47-52.
- Garsia-Sanchez, F., J. L. Jufon, M. Carvaial and J. P. Syverstem. 2002. Gas exchange, chlorophyll and nutrient contents in relation to Na+ and Cl- accumulation in 'Sunburst' mandarin grafted on different rootstocks. Plant Sci., 162: 705-12.
- Ghai N., R. C. Setia and N. Setia. 2002. Effect of paclobutrazol and salicylic acid on chlorophyll content, hill activity and yield components in *Brassica napus* L. (cv. GSL-1). Phytomorphology, 52: 83–87.
- Greenway, H. and R. Munns. 1980. Mechanism of salt tolerance in non-halophytes. Ann. Rev. Plant Physiol., 31: 149-190.
- Hayat, S. and A. Ahmad. (eds) 2007. Salicylic Acid: A Plant Hormone. Springer Publ., Dordrecht.
- Hayat, S., Q. Hayat, M. N. Alyemeni, A. S. Wani, J. Pichtel and A. Ahmad. 2012. Role of proline under changing environments. Plant signal and Behav. Nov. 1: 7(11): 1458-1466.
- Hendawy, S. F. and K. H. A. Khalid. 2005. Response of sage (*Salvia officinalis* L.) plants to zinc application under different salinity levels. J. Appl. Sci. Res., 1: 147-55.
- Hong, Z., K. Lakkinel, Z. Zhang and D. P. S. Verma. 2000. Removal of feedback inhibition of pyrroline-5-carboxylase synthetase results in increased proline accumulation and protection of plants from osmotic stress. Plant Physiol., 122: 1129-1136.
- Jamil, M., D. B. Lee, K. Y. Jung, S. C. Lee and E. S. Rha. 2006. Effect of salt (NaCl) stress on germination and early seedling growth of four vegetables species. J. Central. Euro. Agric., 7(2): 273-282.

- Janković, M. 1974. Genus *Mentha* L. In: M. Josifović (Ed.). Flora of SR Serbia. 6: 524-525.
- Jenifer, A. and J. Franklin- Janus. 2002. Growth and elemental composition of jak pine seedlings treated with sodium chloride and sodium sulfate. Trees structure and function., 16(4- 5): 325-330.
- Kato, M., and S. Shimizu. 1985. Chlorophyll metabolism in higher plants. VI. Involvement of peroxidase in chlorophyll degeneration. Plant Cell Physiol., 26(7): 1291-1301.
- Khan, M. A. and N. C. Duke. 2001. Halophytes- A resource for the future. Wetland Ecology and Management., 6: 455-456.
- Khodary S. E. 2004. Effect of salicylic acid on the growth, photosynthesis and carbohydrate metabolism in saltstressed maize plants. Int. J. Agric. and Biol., 6: 5–8.
- Kim, H., F. Chen, X. Wang and N. C. Rajapakse. 2006. Effect of methyl jasmonate on secondary metabolites of sweet basil (*Ocimum basilicum* L.), J. Agric. Food Chem., 54(6): 2327-32.
- Komali, A. S. and K. Shetty. 1998. Comparison of the growth pattern and rosmarinic acid production in rosemary shoots and genetically transformed callus cultures. Food Biotech., 12(12): 27-41.
- Koocheki, A., M. Nassiri-Mahallati and G. Azizi. 2008. Effect of drought, salinity, and defoliation on growth characteristics of some medicinal plants of Iran. J. Herbs Spices Med. Plants., 14(1-2): 37-53.
- López-Arnaldos, T., M. López-Serrano, A. Ros Baceló, A. A. Calderón and J. M. Zapata. 1995. Spectrophotometric determination of rosmarinic acid in plant cell cultures by complexation with Fe2+ ions. Anal. Chem., 351(2): 311-314.
- Mansour, M. M. F. 2000. Nitrogen containing compounds and adaptation of plants to salinity stress. Biol. Plant., 43: 491-500.
- Marcelis, L. F. M. and J. V. Hooijdonk. 1999. Effect of salinity on growth, water use and nutrient use in radish (*Raphanus sativus* L.). Plant Soil., 215(1): 57-64.
- Mehta, J., R. Naruka, M. Sain, A. Dwivedi, D. Sharma and J. Mirza. 2012. An efficient protocol for clonal micropropagation of *Mentha piperita* L. (Pipperment) Asian J. Plant Sci. and Res., 2(4): 518-523.
- Molsaghi, M., A. Moieni and D. Kahrizi. 2014. Efficient protocol for rapid *Aloe vera* micropropagation. Pharm. Biol., 52(6):735-739.
- Moran, R. and D. Porath. 1980. Chlorophyll determination in intact tissues using *N*,*N*-dimethylformamide. Plant Physiol., 65: 478-479.
- Munjal, R. and C. L. Goswami. 1995. Response of chloroplastic pigments to NaCl and GA3 during cotton cotyledonary leaf growth and maturity. Agric. Sci. Digest., 15: 146-150.
- Muthukumarasamy, M., D. S. Gupta and R. Panneerselvam. 2000. Influence of triadimefon on the metabolism of NaCl stressed radish. Biol Plant., 43: 67-72.
- Naghibi, F., M. Mosaddegh, S. M. Motamed and A. Ghorbani. 2005. Labiatae family in folk medicine in Iran: From ethnobotany to pharmacology. Iran. J. Pharm. Res., 4: 63-79.

- Najafi, F., R. A. Khavari-Nejad and M. S. Ali. 2010. The effects of salt stress on certain physiological parameters in summer savory (*Satureja hortensis* L.) plants. J. Stress Physiol. Bioch., 6(1): 13-21.
- Namdeo, A. G., T. S. Jadhav, P. K. Rai, S. Gavali and K. R. Mahadik. 2007. Precursor feeding for enhanced production of secondary metabolites. Pharm. Rev., 1(2): 227-231.
- Nassiri Asl, M. and H. Hosseinzadeh. 2008. Review of pharmacological effects of *Glycyrrhiza sp* and its bioactive compounds. Phyto. Res., 22: 709-724.
- Nikolova, M. T. and S. V. Ivancheva. 2005. Quantitative flavonoid variations of *Artemisia vulgaris* L. and *Veronica chamaedrys* L. in relation to altitude and polluted environment. Acta. Biol. Szegediensis., 49(3-4): 29-32.
- Osman, M. E. H., S. S. Elfeky, K. Abo El-Soud and A. M. Hasan. 2007. Response of *Catharanthus roseus* shoots to salinity and drought in relation to vincristine alkaloid content. Asian. J. Plant. Sci., 6: 1223-8.
- Phatak, S. V. and M. R. Heble. 2002. Rosmarinic acid synthesis in shoot cultures of *Mentha arvensis* Linn. Ind. J. Biotechl., 1(4): 381-385.
- Queslati, S., N. Karray-Bouraoui, H. Attia, M. Rabhi, R. Ksouri and M. Lachaal. 2010. Physiological and antioxidant responses of *Mentha pulegium* (Pennyroyal) to salt stress. Acta. Physiol. Plant., 32(2): 289-96.
- Razzaque, A. and B. E. Ellis. 1977. Rosmarinic acid production in *Coleus* cell cultures. Planta., 137: 287-291.
- Roy, D. and S. Mukhopadhyay. 2012. Enhanced rosmarinic acid production in cultured plants of two species of *Mentha*. Ind. J. Exp. Biol., 50: 817-825.
- Saliem, A. 2000. Capacity for proline accumulation during salt-stress in *Vicia faba* L. Crop Sci., 17: 90-99.
- SAS Institute, Inc. 2007. SAS Technical Report AS/STAT Software: Changes and Enhancements Users Guide Volume 2, Version 9. 1. 3, Fourth Edition, Cary, NC: SAS Institute, Inc.

- Shao, H. B., L. Y. Chu and C. A. Jaleel. 2009. Water-deficit stress-induced anatomical changes in higher plants. C. R. Biologies., 331(3): 215-25.
- Šiler, B., D. Mišić, B. Filipović, Z. Popović1, T. Cvetić and A. Mijović. 2007. Effects of salinity on *in vitro* growth and photosynthesis of common centaury (*Centaurium erythraea* Rafn.). Arch. Biol. Sci., Belgrade. 59 (2): 129-134.
- Soussi, M., A. Ocana and C. Liuch. 1998. Effects of salt stress on growth, photosynthesis and nitrogen fixation in chickpea (*Cicer arietinum* L.). J. Exp. Bot., 49: 1329-1337.
- Stamenković, V. 2005. Our Non-harming Medicinal Herbs, 2nd revised and expanded edition, Serbia.
- Steel, R. G. and J. H. Torrie. 1980. Principles and Procedures of Statistics 2nd ed. McGraw Hill, NewYork, U.S.A.
- Szabo, E., A. Thelen and M. Peterson. 1999. Fungal elicitor preparations and methyl jasmonate enhance rosmarinic acid accumulation in suspension cultures of *Coleus blumei*, Plant Cell Reports., 18: 485-489.
- Thorpe, T. A. 2007. History of plant tissue culture. Mol Biotechnol. 37: 169–180.
- Weimberg, R. 1975. Effect of growth in highly salinized media on the enzymes of the photosynthetic appartures in pea seedlings. Plant. Physiol., 6: 8-12
- Xu, X., H. Xu, Y. Wang, X. Wang, Y.Qiu and Bo. Xu. 2008. The effect of salt stress on the chlorophyll level of the main sand - binding plants in the shelterbelt along the Tarim Desert Highway. Chin .Sci. Bul., 53: 109-111.
- Yang, F., X. Xiao, Sh. Zhang and Ch. Li. 2009. Salt stress responses in *Populous cathagana* Rehder. Plant. Sci., 176: 669-677.
- Zaki, M. F., S. D. Abou-Hussein, M. M. Abou El-Magd and H. M. H. El-Abagy. 2009. Evaluation of some sweet fennel cultivars under saline irrigation water. Eur. J. Sci. Res., 30(1): 67-78.

الملخص العربى

تأثير كلوريد الصوديوم والفينيل الآنين على إنتاج بعض المركبات الثانوية المُنتجَة معملياً في نبات الثير كلوريد الصوديوم والفينيل الأنعناع طويل الأوراق

علا عبد العزيز الشناوي، محمد جمال التركي، هدى السيد المقدم، باسم إبراهيم عباس

زراعة الأوراق استخدم الBA (٥,٠ مليجرام/لتر) وال 2,4-D (٤ مليجرام/لتر) معا، وتم ضبط رقم الحموضة 5.8±0.02. تم تحليل الأفرع الخضرية الناتجة بعد ٦٠ يوم من الزراعة، ونسيج الكالس تم تحليله بعد ٦٠ يوم مرة وبعد ١٠٠ يوم مرة أخرى لتقدير نسبة المركبات الفينولية الكلية وحمض الروزماري والبرولين المتراكمين داخل أنسجة النباتات. تفوقت الأفرع الخضرية في المحتوى الكلى للكلوروفيل والمحتوى الكلى للمواد الفينولية وحمض الروزمارينك مقارنة بمحتواهم في نسيج الكالس، استخدام التركيزات المنخفضة من كلوريد الصوديوم (۲۰۰۰ مليجرام/لتر) لتحفيز إنتاج وتراكم بعض المركبات والمواد الثانوية الفعالة يعتبر الحل الأمثل، مع تجنب التركيزات المرتفعة (٢٠٠٠ مليجرام/ لتر)، بينما التركيزات المتوسطة والمرتفعة من الحامض الأميني فينيل ألانين (٥، ١٠، ١٥ مليجرام/ لتر) تعتبر الحل الأمثل لتحفيز إنتاج وتراكم بعض المركبات الثانوية الفعالة.

أجريت هذه الدراسة بمعمل زراعة الأنسجة بقسم الزهور ونباتات الزينة وتنسيق الحدائق بكلية الزراعة (الشاطبي) جامعة الإسكندرية خلال الفترة ٢٠١٤-٢٠١٥م، بهدف تعزيز إنتاج بعض المركبات الثانوية المتراكمة في أنسجة نبات النعناع طويل الأوراق معملياً تحت تأثير كلوريد الصوديوم والحامض الأميني فينيل ألانين. تم أخذ النباتات والبالغ عمرها شهرين من مشتل تجارى وزراعتها معمليا ووضعها تحت ظروف إضاءة صناعية باستخدام لمبات الفلورسنت ١٦ ساعة إضاءة وعلى درجة حرارة ٢٥°م تم زراعة القمم النامية بطول ٣–٤ سم وزراعة الأوراق اسم٢ على بيئة غذائية معقمة مضاف إليها السكروز ٣٠ جم/لتر، والآجار ٨ جم/لتر، وكلوريد الصوديوم (٠، ٢٠٠٠، ٢٠٠٠، ٢٠٠٠ مليجرام / لتر) وفينيل ألانين (٠، ٥،، ٥، ١٠، ١٥ مليجرام/ لتر)، في حالة زراعة القمم النامية استخدم الNAA (٠,٢٥ مليجرام /لتر)، والBA (٢,٥ مليجرام/لتر) معا، عند