

Potential Protective Effects of Ethanolic Extract of Butterfly Pea (*Clitoria ternatea* Linn) Flower Against Carbon Tetrachloride-Induced Hepatotoxicity in Rats

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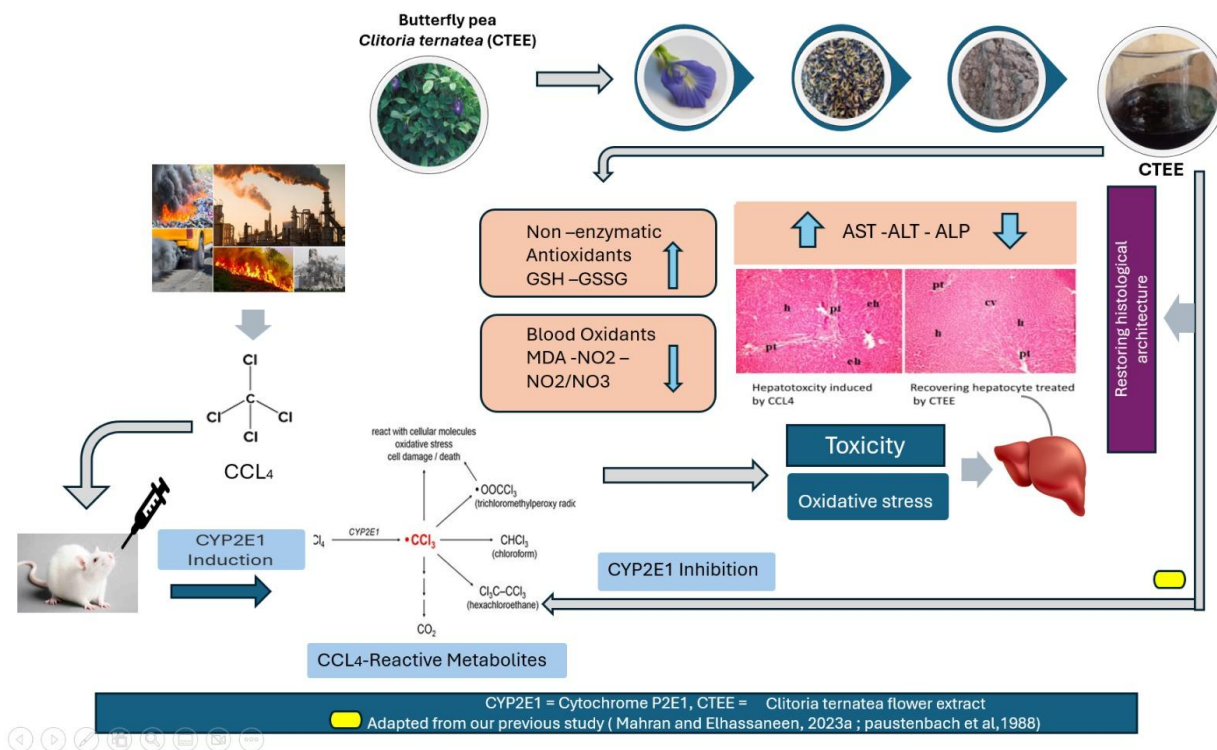
ABSTRACT

The current study was carried out to explore the hepatoprotective properties of ethanol-derived *Clitoria ternatea* flower extract (CTEE) on liver injury induced by carbon tetrachloride in rats. It was observed that rats injected with carbon tetrachloride (CCl₄) exhibited significant (p<0.05) reductions in various biological parameters like BWA, FI, and FER, with percentages of -42.86%, -36.99%, and -30.14%, respectively, compared to rats in the control group. Moreover, in comparison to the rats in the normal group, biochemical parameters like total bilirubin (TB), AST, ALT, ALP, and 129.90 liver enzyme activities showed significant (p<0.05) elevations at 77.28, 129.90, 158.72, and 412.84%, respectively. Along with a decrease in the level of non-enzymatic antioxidants (GSH and GSSG) and an increase in the level of oxidants (MDA and NO₂), this was also correlated with an

imbalance in certain antioxidant parameters and oxidants in the blood. Each of these parameters suggested that CCl₄ was the cause of the liver injury. Rat protocol feeding with CTEE intervention produced a significant (p<0.05) improvements in all previous biological, biochemical, and oxidant/antioxidant status markers, indicating protection against hepatotoxicity. Furthermore, a dose-dependent pattern was seen in the rate of improvement for each of these parameters. In conclusion, CTEE demonstrated efficacy in preventing liver disorders caused by CCl₄. It is therefore advised that we include such plant parts (flowers of *Clitoria ternatea*) in our daily diets, beverages, and pharmaceutical formulae at concentrations of up to 400 mg/kg bw/day.

Keywords: *Clitoria ternatea*, body weight, liver functions, glutathione fractions, malonaldehyde, nitrite.

Graphical abstract



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INTRODUCTION

Originally from equatorial Asia, which includes parts of South and Southeast Asia, *Clitoria ternatea* (family: *Fabaceae*) is also known as butterfly pea. It has been introduced to Africa, Australia, and the Americas. With elliptic, obtuse leaves, it is a perennial herbaceous plant (Patel, 2023). The flower of this plant is its most valuable component because it is used in ayurvedic medicine and as a natural food coloring for desserts and glutinous rice (Vuong and Parichat, 2021). Typically, dried lemongrass and other herbs are added to alternative flowers to make butterfly pea flower tea. When lemon juice turns purple, the mixture's color varies based on what is added to the liquid (Pantazi, 2016). For instance, to boost the acidity and give the beverage a purplish-pink hue, blue butterfly pea flower tea is typically combined with honey and lemon in Thailand and Vietnam. This results in a beverage that is typically served hot or cold after supper.

This plant has been linked to a number of different effects in traditional medicine, including sedative, antidepressant, anticonvulsant, anxiolytic, memory-enhancing, nootropic, and anti-stress properties. In conventional Chinese medicine (Chakraborty *et al.*, 2018). Additionally, studies on plant extracts have demonstrated a strong ability to lessen the severity of behavior induced by acetylcholine and serotonin (Neeti *et al.*, 2003). Additionally, numerous biological and pharmacological activities were observed in its extracts, such as wound healing activity, inhibition of platelet aggregation, antimicrobial, antipyretic, anti-inflammatory, analgesic, diuretic, local anesthetic, anti-diabetic, and its use as vascular smooth muscle relaxing properties (Solanki & Jain, 2012; Nithianantham *et al.*, 2013 and Sreeharini *et al.*, 2013). Consequently, the herb has been used for a very long time in conventional therapies to treat a wide range of illnesses, such as kidney, blood, and diabetes (Pulok *et al.*, 2008). Numerous prior investigations have ascribed the majority of *C. ternatea's* biological effects to a variety of bioactive substances, including flavanol glycosides, anthocyanins, steroids, triterpenoids, and nucleotides, which are cyclic peptides (Mukherjee *et al.*, 2008). Moreover, previous reports (Gupta *et al.*, 2010 and Pendbhaje *et al.*, 2011) described the isolation and identification of phenolics, flavonoids, and different kaempferols from *C. ternatea* flowers. These substances are said to be in charge of their scavenging and antioxidant properties (Kuppan *et al.*, 2013). The presence of multiple anthocyanins, the most significant of which are ternatins, polyacylated derivatives of delphinidin 3,3', 5'-triglucoside, accounts for the blue

color of this flowering plant as well (Vidana Gamage *et al.*, 2021).

The synthesis of the factors that required to prevent blood bleeding, the production of bile to aid in the absorption of nutrient, the processing of numerous products released into the bloodstream (such as glucose, plasma proteins, and urea), and the storage of various products (such as glycogen, fat, vitamins, and minerals) are just a few of the critical roles played by the vital organ, the liver (Kebamo *et al.*, 2015). Furthermore, it is essential for the removal, metabolism, and biotransformation of foreign substances as well as for preserving the biological equilibrium of the human body (Grażyna *et al.*, 2020 and Yu *et al.*, 2020). Accordingly, harm to the liver can result in a variety of conditions, from cirrhosis and potentially fatal liver failure to a brief increase in liver enzymes (Bera *et al.*, 2012). Numerous substances that are frequently used can harm liver cells and metabolism (Meyer and Kulkarni, 2001). Carbon tetrachloride (CCl₄) is the most widely used chemical. Based on studies indicated evidence of carcinogenicity in experimental animals, it is expected to be a human carcinogen. It also has hepatotoxic effects by triggering steatosis and centrilobular necrosis (WHO, 1999). According to reports, the cytochrome P-450 system biotransforms CCl₄ to create trichloroethylene free radicals, which is the mechanism of CCl₄-induced hepatic injury. The trichloroethylene peroxy radicals that are created when these free radicals react with oxygen act on the endoplasmic reticulum's lipid membrane to cause lipid peroxidation (Weber *et al.*, 2003 and Wang *et al.*, 2024). All of those earlier investigations, in addition to others, suggested that CCl₄ is one of the most effective models of liver injury. Liver disease continues to be a global health concern even with the amazing advancements in research. As a result, the ongoing and significant task of finding novel pharmaceuticals was undertaken (Nithianantham *et al.*, 2011). To support these claims, numerous scientific studies and research are required. In this context, plant and natural products have been used in traditional treatments throughout the world to prevent or treat liver diseases. These assertions require a great deal of scientific investigation (Lahon *et al.*, 2011). As far as we know, though, there hasn't been any evidence of the hepatoprotective potential of *C. ternatea* flowers against liver damage caused by carbon tetrachloride in rats. Thus, the goal of the current investigation was to determine whether ethanolic *C. ternatea* flower extract could shield rats' livers from carbon tetrachloride-induced damage.



Fig.1 Whole plant of *Clitoria ternatea*



Fig. 2 *C. ternatea* L. The Morphoforms: A, Single form



Clitoria ternatea L. (Flowers)



Clitoria ternatea L. (Powder)

Botanical description of a plant

Clitoria ternatea displays twining fine stems that can reach a length of 0.5-3 m. The leaves exhibit a pinnate arrangement, consisting of 5-7 elliptic to lanceolate leaflets measuring 3-5 cm in length and featuring a short pubescence underneath (Fig. 1). The flowers are solitary, showcasing hues of deep blue to blue mauve, with very short pedicels and a length of 4-5 cm (Fig. 2). Its pods are beaked flat, linear, 5-10 cm long, 0.6-1.4 mm wide, and mildly pubescent, containing up to 10 seeds. These seeds are olive and brown in color, often displaying a mottled pattern, measuring 4.0-8 mm in length and 3.2-4.5 mm in width (Mehmood *et al.*,2019; Sarma *et al.*,2023 and Arya *et al.*,2022).

MATERIALS AND METHODS

Materials

The dried flowers of *Clitoria ternatea* Linn, commonly known as butterfly pea, were procured from Bab ElKhalk, Cairo, Egypt's El Misryia Company for Trading Herbs and Medical Plants. Taxonomic verification of *Clitoria ternatea* was conducted by the University of Alexandria's Faculty of Agriculture's Agricultural Plant Department, which is situated in Alexandria, Egypt. Casein was sourced from Morgan Chemical Co. in Cairo, Egypt. Carbon tetrachloride (CCl_4), in the form of a 10% liquid solution, along with all necessary chemicals, buffers, and solvents, were supplied by ElGhohorya Company for Trading Drugs, Chemicals, and Medical Instruments in Cairo, Egypt. The assays included in the kit for assessing Alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), and malondialdehyde (MDA) were acquired from (BIODIAGNOSTIC in Dokki). Additionally, albumin (Alb) assays were obtained from (El-Nasr

Pharmaceutical Chemicals Company), while (GSH, and GSSG) assays were procured from (Nawah Scientific in Almokattam, Cairo, Egypt).

Methods

Preparation of *Clitoria ternatea* flowers ethanol extract (CTEE)

Parimaladevi *et al.*,(2003) and Elhassaneen *et al.*, (2023) instructions were followed in the preparation of the *Clitoria ternatea* flowers ethanol extract (CTEE). To put it briefly, samples of dried flowers from *Clitoria ternatea* were brought into the lab. After being manually removed, the flowers were sieved to remove any foreign objects. The flowers were ground into a powder (20 mesh) and subsequently combined following grinding at a high speed using a miller (Moulinex Egypt, Al-Araby Co., Egypt) to produce uniform samples. A quantity of twenty grams of dried *Clitoria ternatea* powder was subjected to extraction with 80% ethanol for a duration of 5–6 hours (20 ± 3 minutes per cycle) utilizing a Soxhlet apparatus. The final step involved the evaporation of the solvent under reduced pressure in order to extract the dry solvent, a process carried out using a (rotary evaporator) (**Buchi R-210, Switzerland**). Subsequently, the extract was stored at a temperature of 4 °C until required. The total yield of *Clitoria ternatea* ethanol extract (CTEE) was determined to be 5.61% (w/w) in terms of yield.

Proximate analysis of *Clitoria ternatea* flower extract (CTEE)

The method described by Official Agricultural Chemists (AOAC) International (1990) was used to determine Proximate Analysis. Moisture, ash, fat, protein concentration, fiber, and carbohydrates are also included in the six analyses.

Biological (in-vivo) experiments

Ethical approval

Biological experiments conducted for this research were granted ethical approval by (The Institutional Animal Care and Use Committee (IACUC) at Alexandria University, Egypt). Approval ID/number for this study is ALEXU/.011/ 2023.

Animals

The study utilized animals, specifically adult male albino rats (180±6.4 g each), procured from (The Research Institute of Ophthalmology, Department of Medical Analysis, located in Giza, Egypt). The formulation of the Basal Diet (BD) includes (10%) Protein, (10%) corn oil, (1%) vitamin mixture, (4%) mineral mixture, (0.2%) choline chloride, (0.3%) methionine, (5%) cellulose, and corn starch (69.5%), as outlined by AIN (1993). The same prior reference was used to formulate the components of vitamin and salt mixtures.

Induction of hepatotoxic rats

Using the technique developed by Khan *et al.*, (2012), Forty-two (Albino rats, male) were administered intraperitoneal (IP) injections of tetrachlorocarbon (CCl₄) diluted in olive oil at a concentration of (50% V/V)-(2 ml/kg bw) twice weekly for a duration of two-week to induce chronic liver injury. A biochemical analysis of three rats chosen at random from the experimental group of animals and

confirmed with histological study under light microscopy illustrated the presence of liver damage.

Experimental design

The Commission on Life Sciences, the National Research Council, and the Institute of Laboratory Animal Resources all issued guidelines that were adhered to in biological experiments (NRC, 1996). A total of 42 rats were housed individually in wire cages under standard conditions with a 12-hour light-dark cycle, temperature maintained at 25 ± 2°C, and relative humidity at 52 ± 3%. Before commencing the study, all rats were fed BD for a two-weeks acclimatization period. Subsequently, the rats were randomly allocated into two primary groups. The initial group of six rats served as the negative control (normal) and received BD as food along with a paraffin oil injection (5 ml/kg bw) as a vehicle for CCl₄ treatment. The second main group comprised (36) rats injected with CCl₄ to induce hepatotoxicity; these rats were further segregated into sex-matched subgroups (Groups 3–7) and administered CTEE orally at doses of (25, 50, 100, 200, and 400) mg/kg bw/day. A positive control group (model) receiving BD was assigned as group 2. Each group was accommodated in a single cage for 28 days, as depicted in Table (1). The selection of CTEE concentrations was guided by acute toxicity studies conducted by Sreerhini *et al.*, (2013), which demonstrated the safety of CTEE up to 5000 mg/kg bw via oral administration. Rat weights were recorded at the beginning of the experiment, weekly thereafter, and upon the conclusion of the study period.

Table 1. Experimental design to assess the potential protective effects of CTEE against CCl₄ induced hepatotoxicity in rats

Groups	Interventions	
Group I Negative control (- Ve)	Normal control	BD for 28-day
Group II Positive control (+Ve)	CCl ₄ control	BD +5 ml/kg bw paraffin oil injection for 28-day
Group III	CCl ₄ + CTEE 25 mg/kg BW/day)	2 mg/kg/BW of CCl ₄ in olive oil at a 50% V/V for 28-day+ 25 mg/kg BW/day of CTEE
Group IV	CCl ₄ + CTEE 50 mg/kg BW/day)	2 mg/kg/BW of CCl ₄ in olive oil at a 50% V/V for 28-day+ 50 mg/kg BW/day of CTEE
Group V	CCl ₄ + CTEE 100 mg/kg BW/day)	2 mg/kg/BW of CCl ₄ in olive oil at a 50% V/V for 28-day+ 100 mg/kg BW/day of CTEE
Group VI	CCl ₄ + CTEE 200 mg/kg BW/day)	2 mg/kg/BW of CCl ₄ in olive oil at a 50% V/V for 28-day+ 200mg/kg BW/day of CTEE
Group VII	CCl ₄ + CTEE 400 mg/kg BW/day)	2 mg/kg/BW of CCl ₄ in olive oil at a 50% V/V for 28-day+ 400 mg/kg BW/day of CTEE

* CCl₄ : carbon tetrachloride; CTEE: *Clitoria tematea* flower extract

Biological evaluation

Every day of the 28-day experiment was monitored for diet consumption, and every week, body weight was recorded. In accordance with Chapman *et al.*, (1959), calculations were made for weight body gain (BWG%), intake food (FI), and efficiency food ratio (FER) using specific equations: BWG (%) = (Final weight - initial weight) / initial weight × 100, and FER (grams) = feed intake (grams/28 days) / grams of body weight gain.

Sampling collection

Rats underwent a 12-hour fasting period before blood samples were collected at the conclusion of the 28-day experimental period. Subsequently, the rats were euthanized under ether anesthesia and dissected.

Blood sampling

Following established procedures (Parasuraman *et al.*, 2010 and Drury & Wallington, 1980), blood samples collected from the abdominal aorta were transferred to sterile, dry centrifuge tubes. The samples were kept at room temperature to clot (25°C) before centrifugation for 10 minutes at 3500 rpm to separate the serum. After centrifugation, the serum was carefully aspirated and transferred to sterile capped tubes for storage at -80°C until further analysis.

Liver sampling

Following blood collection, the rats were rendered unconscious during blood sampling, and their liver was immediately removed, perfused with ice-cold 0.9% normal saline, dried, and weighed using an electronic balance. Liver organ was extracted, and the relative organ weight (percentage of body weight) was calculated according to Mee-Young *et al.*, (2013)'s formula. The liver organ was rinsed in 10% neutral formalin for preparing paraffin sections to study the histopathological changes under light microscopy.

Hematological Biochemical analysis

Liver functions

Being serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST) levels quantified in serum in the assay implemented by Gella *et al.*, (1985) and Tietz *et al.*, (1994), the instrumental solution was adopted for the accomplishment of the assay. Also, entrusted with the kinetic activity of ALP was done with the modified method invented by Vassault *et al.*, (1999).

Biological - Antioxidant enzymes

The concentrations of reduced glutathione (GSH) and oxidized glutathione (GSSG) were determined in serum samples using established colorimetric methods, as outlined by Ellman (1959).

Biological oxidants

Using a fluorometric method similar to that detailed by Misko *et al.*, (1993), serum nitrite (NO₂) was measured. The method of thiobarbituric acid (TBA) is used to measure the amount of serum malondialdehyde (MDA), as described by Buege and Aust (1978).

Histopathological technique

The liver fixed in 70%, 80%, 95%, and absolute alcohol for dehydration, followed by clearing in xylene. Subsequently, the liver tissue underwent impregnation in molten wax to create a paraffin block. These paraffin blocks were then sliced into 5µm thick sections, which were placed on pristine slides and incubated in a 37°C oven. The sections were deparaffinized in xylene, rehydrated gradually in alcohol, and finally stained with Haematoxylin and Eosin. Post-staining, they were dehydrated in increasing alcohol concentrations and cleared in xylene. Following this, the slides were covered with coverslips, examined, and photographed under light microscopy at the Histochemistry and Cell Biology department of the Medical Research Institute, Alex. Univ.

Statistical Analysis

A computerized Computer Software Package SPSS® version (18.0.) program was used to statistically analyze all the data using one-way ANOVA. The means ± Standard Deviation (SD) were reported for the results. P ≤ 0.05 was used to determine the significance of differences between treatments (Snedecor and Cochran, 1967).

RESULTS AND DISCUSSION

Proximate analysis of *Clitoria ternatea* flower extract (CTEE)

Table (2) indicated the results of the proximate analysis including moisture content, crude protein, ash, crude fat, and crude fiber. Moisture 93.04, Crude protein 0.35, Ash 0.42, Fat 2.53, Crude fiber 2.21, Carbohydrate 1.45. There was an obvious quantity of fat (2.53 ± 0.1) and crude fiber (2.21 ± 0.2) in the *C. ternatea* flowers. The results agree with Neda *et al.*, (2013) and Muhammad Ezzudin & Rabeta (2018) who found that higher protein content in the leaves and flower of *C. ternatea* could be useful against muscle wasting.

Table 2. Proximate analysis of *Clitoria ternatea* flower extract (CTEE)

Constituent	Percentage (%)
Moisture	93.04 ±0.22
Protein	0.35 ±0.17
Ash	0.42 ±0.11
Fat	2.53±0.09
Crude fiber	2.21±0.04
Carbohydrate	1.45 ±0.08

Biological evaluation

Effect of intervention with (CTEE on BWG, FI and FER) of hepatotoxic rats induced by (CCl₄)

The influence of CTEE interfere (BWG, FI, and FER) on rats subjected to CCl₄-induced hepatotoxicity is presented in Table (3). Upon analysis of this data, it was noted that rats treated with CCl₄ exhibited a notable decrease at ($p \leq 0.05$) in BWG (-42.86%), FI (-36.99), and FER (-30.14) compared to the control group. Conversely, in rats undergoing a 28-day feeding regimen, the administration of CTEE at varying doses of 25, 50, 100, 200, and 400 mg/kg bw/day a

significantly ($p \leq 0.05$) elevated the levels of BWG, FI, and FER by -40.66, -34.07, -16.48, -10.99, and -8.79%; -30.41, -25.47, -16.30, -8.62, and -6.41%; and -27.40, -20.55, -15.03, -12.49, and -6.90%, respectively, in comparison to the normal animals.

Furthermore, there was a dose-dependent pattern to the rate of increase in each of these parameters. In related investigations, multiple researchers discovered that CCl₄ significantly reduced the rats' BWG, FI, and FER, and that feeding interventions containing various plant parts significantly increased these parameters as well (ElSamouny, 2021 and Elhassaneen *et al.*, 2021 a,b). According to Hamzawy *et al.*, (2013) and Abd El-Rahman (2021), hepatotoxic rats showed a notable decrease in body weight and feed intake in this regard. Additionally, Ross *et al.*, (2020) and Weber *et al.*, (2003) demonstrated that hepatotoxic treatment can cause malnutrition, characterized by inadequate dietary intake (FI), impaired digestion, and subsequent malabsorption of nutrients. Additionally, this condition may involve anomalies in the storage and metabolism of nearly all nutrients, which are the main causes of malnutrition in hepatotoxic patients. Additionally, Boushey *et al.*, (2001) and Khan *et al.*, (2012) noted that the majority of hepatotoxic patients lose weight and are sick.

Table 3. Effect of CTEE intervention on (BWG., FI., and FER.) of hepatotoxic rats induced by CCl₄

Value	Normal cont.	Model cont.	CTEE intervention (mg/kg bw/day)				
			25	50	100	200	400
BWG (%)							
Mean	0.94 ^a	0.54 ^d	0.56 ^d	0.62 ^c	0.78 ^{bc}	0.83 ^b	0.85 ^b
SD	0.03	0.05	0.03	0.04	0.09	0.04	0.05
% of change	0.00	-42.86	-40.66	-34.07	-16.48	-10.99	-8.79
FI (g/day/rat)							
Mean	13.14 ^a	8.28 ^d	9.15 ^c	9.80 ^c	11.00 ^{bc}	12.01 ^b	12.30 ^b
SD	0.31	0.42	0.51	0.62	0.70	0.51	0.49
% of change	0.00	-36.99	-30.41	-25.47	-16.30	-8.62	-6.41
FER							
Mean	0.075 ^a	0.053 ^d	0.055 ^d	0.060 ^{cd}	0.064 ^c	0.066 ^{bc}	0.070 ^b
SD	0.004	0.006	0.003	0.005	0.003	0.006	0.01
% of change	0.00	-30.14	-27.40	-20.55	-15.03	-12.49	-6.90

Each value represents the mean ±SD (n = 6). Means in the same column with different superscript letters indicate the significant differences at $P \geq 0.05$. Natural control, healthy rats with no intervention; form control, carbon tetrachloride induced hepatotoxic rats without intervention; CTEE, *Clitoria ternatea* flower ethanolic extract; CTEE intervention, carbon tetrachloride-induced hepatotoxic rats with CTEE intervention; bw, body weight; BWG, body weight gain; FI, feed intake; FER, feed efficiency rate.

At last, a number of authors discovered that rats given CCl₄ injections showed decreased BWG, FI, and FER, and that these levels could be raised by consuming or interacting with various plant parts that contain bioactive compounds, like those in CTEE (Mansour, 2017; Tahoon, 2019; ElSamouny, 2021 and Elhassanen *et al.*, 2021 a,b).

Biochemical Finding

Effect of intervention with CTEE on the liver functions of the hepatotoxic rats induced by CCl₄

Table (4) illustrates the effects of CTEE intervention on a hepatic function of CCl₄-induced hepatotoxic rats. comparing the CCl₄-treated rats companion to the control group, the results indicated a substantial elevation ($p \leq 0.05$) in AST (77.28%), ALT (129.90%), ALP (158.72%), and TB (412.84%). However, following CTEE intervention (25, 50, 100, 200, and 400 mg/kg bw/day) in rats subjected to a 28-day dietary regimen, the levels of AST, ALT, ALP, and TB

exhibited a significant decline compared to the control animals ($p \leq 0.05$). The reductions were recorded as 68.62, 52.80, 42.21, 23.25, and 16.25%; 129.51, 110.88, 97.28, 67.45, and 28.17%; 153.00, 141.95, 119.53, 77.92, and 42.40%; and 366.22, 306.76, 239.86, 177.70, and 64.86%, respectively. Furthermore, a dose-dependent relationship was observed in the rate of decrease for each of these parameters. According to several researchers (Elhassanneen *et al.*, 2012 and Susilo *et al.*, 2019), CCl₄ - induced liver injury is regularly employed to investigate the hepatoprotective effects of pharmaceuticals and natural substances. CCl₄ is metabolized by liver cytochrome P450 into trichloromethyl (CCl₃-) radicals, which then combine with O₂ to create trichloromethyl peroxy (CCl₃OO-) radicals, leading to liver damage. These free radicals disrupt the formation and purpose of intracellular organelles and cell membranes in liver cells, causing the peroxidation of membrane-bound fatty acids.

Table 4. Effect of intervention with CTEE on the liver functions of the hepatotoxic rats induced by CCl₄

Value	Normal cont.	Model cont.	CTEE intervention (mg/kg bw/day)				
			25	50	100	200	400
Serum Aspartate aminotransferase (AST) activity (U/L)							
Mean	38.58 ^c	68.39 ^a	65.05 ^a	58.95 ^b	54.86 ^b	47.55 ^c	44.85 ^c
SD	3.88	5.13	4.60	6.03	4.40	3.84	5.26
% of change	0.00	77.28	68.62	52.80	42.21	23.25	16.25
Serum alanine aminotransferase (ALT) activity (U/L)							
Mean	26.78 ^e	61.57 ^a	61.47 ^a	56.48 ^{ab}	52.83 ^b	44.84 ^c	34.33 ^d
SD	2.26	6.34	3.99	3.76	4.12	2.99	3.45
% of change	0.00	129.90	129.51	110.88	97.28	67.45	28.17
Serum alkaline phosphatase (ALP, U/L)							
Mean	100.96 ^e	261.19 ^a	255.41 ^a	244.26 ^{ab}	221.63 ^b	179.62 ^c	143.76 ^d
SD	9.32	19.14	13.66	15.85	9.10	11.32	11.09
% of change	0.00	158.72	153.00	141.95	119.53	77.92	42.40
Bilirubin (TB, mg/L)							
Mean	1.48 ^e	7.59 ^a	6.90 ^{ab}	6.02 ^b	5.03 ^{bc}	4.11 ^c	2.44 ^d
SD	0.23	0.62	0.41	0.38	0.46	0.29	0.21
% of change	0.00	412.84	366.22	306.76	239.86	177.70	64.86

Each value represents mean \pm SD (n=6). Means on the same column with different superscript letters explain the significant difference at $P \leq 0.05$. Normal control, healthy rats without intervention; Model control, carbon tetrachloride-induced hepatotoxic rats without intervention; CTEE, *Clitoria ternatea* flower ethanolic extract; CTEE intervention, carbon tetrachloride-induced hepatotoxic rats with CTEE intervention.

The elevated levels of serum markers AST, ALT, ALP, and TB in this study indicate significant liver damage induced by CCl_4 injection. According to Shanmugasundaran *et al.*, (2010) and Nithianantham *et al.*, (2011), these variable assays are frequently regarded as sensitive markers for assessing the progression of liver damage since they contain cytoplasm, which promotes blood flow following liver cell damage. According to data from a recent study, CTEE feeding intervention rats' levels of aminotransferases (AST and ALT), ALP, and TB were significantly decreased, demonstrating that it could prevent cell damage. The CTEE content of a number of significant bioactive substances, such as steroids, anthocyanins, flavonol glycosides, triterpenoids, and nucleotides cyclic peptides may be responsible for these protective effects (Mukherjee *et al.*, 2008 and Nithianantham *et al.*, 2011). Moreover, earlier reports on CTEE included phenolics, flavonoids, and different kaempferols (Gupta *et al.*, 2010 and Pendbhaje *et al.*, 2011). According to reports, these substances have antioxidant and scavenging properties, and they play a protective role against toxins like CCl_4 that can cause liver damage (Kuppan *et al.*, 2013; Elhassaneen *et al.*, 2016 and Elhassannen *et al.*, 2021 a,b).

Effect of intervention with CTEE on glutathione fractions of hepatotoxic rats induced by -CCl_4

Table (5) demonstrates the impact of CTEE intervention on the levels of biological antioxidants, specifically glutathione (GSH) fractions, in rats with hepatotoxicity induced by CCl_4 . The analysis allowed for the determination of mean GSH and GSSG values in the normal animal group were 8.98 ± 0.38 and $0.697 \pm 0.067 \mu\text{mol/L}$, respectively. A notable reduction in GSH (-45.43%) and GSSG (-18.16%) levels was observed in the CCl_4 -treated rats when compared to the normal group ($p \leq 0.05$). Conversely, rats that were administered a diet containing different doses of CTEE (25, 50, 100, 200, and 400 mg/kg bw/day) for a duration of 28 days exhibited a significant increase ($p \leq 0.05$) in GSH and GSSG levels by -37.31, -30.03, -23.39, -14.37, and -8.13%, and -19.10, -13.63, -9.08, -8.20, and -2.56%, respectively. Furthermore, a dose-dependent pattern was observed in the rate of increase for each of these parameters. Considerable focus has been placed on the biosynthesis, regulation, and intracellular functions of the tripeptide-thiol glutathione, also known as α -glutamyl cysteinyl-glycine.

Table 5. Effect of intervention with CTEE on glutathione fractions of the hepatotoxic rats induced by CCl_4

Value	Normal cont.	Model cont.	CTEE intervention (mg/kg bw/day)				
			25	50	100	200	400
Reduced glutathione concentration, (GSH, $\mu\text{mol/L}$)							
Mean	8.98 ^a	4.90 ^c	5.63 ^c	6.28 ^{bc}	6.88 ^b	7.69 ^{ab}	8.25 ^a
SD	0.38	0.54	0.55	0.61	0.39	0.48	0.60
% of change	0.00	-45.43	-37.31	-30.03	-23.39	-14.37	-8.13
Oxidized glutathione concentration (GSSG, $\mu\text{mol/L}$)							
Mean	0.697 ^a	0.570 ^c	0.564 ^c	0.602 ^c	0.634 ^b	0.640 ^b	0.679 ^a
SD	0.067	0.094	0.033	0.042	0.069	0.122	0.044
% of change	0.00	-18.16	-19.10	-13.63	-9.08	-8.20	-2.56
GSH/GSSG ratio (%)							
Value	12.886 ^a	8.591 ^d	9.987 ^c	10.439 ^b	10.858 ^b	12.021 ^a	12.149 ^a

Each value represents mean \pm SD (n=6). Means on the same column with different superscript letters explain the significant difference at $P \leq 0.05$. Normal control, healthy rats without intervention; Model control, carbon tetrachloride-induced hepatotoxic rats without intervention; CTEE, *Clitoria ternatea* flower ethanolic extract; CTEE intervention, carbon tetrachloride-induced hepatotoxic rats with CTEE intervention.

The primary function among these is the detoxification process, which is associated with phase II free radical metabolites that are subsequently eliminated in the urine as a result of drug use or chemical pollution (Elhassaneen, 1996 and Rao *et al.*, 2013). Moreover, GSH has super antioxidant properties such as being a nonenzymatic oxyradical scavenger and playing a part in the activation of various antioxidant enzymatic systems, including GSH-Px and GSH-Rd (Almaadawy *et al.*, 2016). The transfer of GSH fractions from the liver to the blood may be impeded by CCl₄, in accordance with a number of earlier authors, including the current study (Elhassaneen *et al.*, 2012 and Abd Elalal *et al.*, 2022). These conditions are attributed to lipid peroxidation, hepatocyte dysfunction, and/or decreased energy due to a decrease in liver glycogen content.

The bioactive substances of CTEE, including anthocyanins, flavonoids, triterpenoids, phenolics, and steroids, exhibit antioxidant properties and enhance the liver's release of GSH into the bloodstream (Mukherjee *et al.*, 2008; Nithianantham *et al.*, 2011 and Mahran & Elhassaneen, 2023). The results of this investigation also showed that a decline in the ratio of GSH/GSSG was observed in conjunction with a decrease in serum GSH fractions. GSH/GSSG ratios in normal liver cells are usually very high, i.e., >10 (Thilavech *et al.*, 2021). The rats given CCl₄ injection showed a significant ($p \leq 0.05$) decrease in their serum redox state (GSH/GSSG). However, after receiving CTEE for up to 400 mg/kg bw/day, the GSH/GSSG ratio increased and approached the value of the rats in the normal group. Therefore, it is suggested that CTEE suppress oxyradical fluxes induced by CCl₄, which could cause the GSH/GSSG ratio to decrease.

Effect of intervention with CTEE on oxidative stress of the hepatotoxic rats induced by- CCl₄

Table (6) illustrates the impact of CTEE intervention on oxidative stress, malondialdehyde (MDA), and nitrite (NO₂) in hepatotoxic rats induced by CCl₄. From these data, it was attainable to determine that the normal animal group's mean MDA and NO₂ values were 0.401 ± 0.024 and 4.95 ± 0.59 nmol/mL, respectively. When contrasted to the normal group, the CCl₄-treated rates showed a substantial ($p \leq 0.05$) increase in MDA (112.17%) and NO₂ (13.36%). However, after 28 days of feeding rats, CTEE intervention (25, 50, 100, 200, and 400 mg/kg bw/day) significantly ($p \leq 0.05$) reduced MDA and NO₂ levels relative to normal animals by 106.35, 96.30, 77.25, 40.74, and 16.40%, and 109.05, 87.93, 54.74, 33.62, and 21.55%, respectively. Furthermore, the rate of decline in each of the aforementioned variables showed a dose-dependent

pattern, as Weber *et al.*, (2003) and Wang *et al.*, (2024) explained. CCl₄ causes liver damage by being biotransformed by the liver cytochrome P-450 into free radicals (trichloromethyl, CCl₃·, and trichloromethyl peroxy (CCl₃OO·). All fatty acids that are bound to membranes experience lipid peroxidation as a consequence of these radicals. Malondialdehyde (MDA), a hazardous degradative product, can form in cell membranes as a result of lipid peroxidation (Cheeseman and Salter, 1993). Due to its association with nitrogenous bases in DNA, MDA has been shown in several studies to have mutagenic impacts (Cline *et al.*, 2004 and Mahran & Elhassaneen, 2023). Moreover, MDA forms a cross-linking with the components of the membrane, which disrupts its fluidity, osmolality, and receptor inactivation. According to Nilanjana (2013), each of these conditions has the potential to damage cells and result in the formation of atherosclerotic plaques. Inversely, L-arginine is enzymatically transformed into citrulline and the biologically active free radical nitric oxide (NO) by nitric oxide synthase in an inverse manner (Manahan, 1989). The detrimental effects of NO can manifest through multiple pathways: 1) conversion into nitrite (NO₂) and nitrate (NO₃) upon interaction with oxygen and water; 2) formation of nitrosylated compounds through interaction with amino and thiol groups of proteins; 3) reaction with hemoglobin leading to the creation of iron-nitrosyl adducts in the bloodstream; and 4) combination with superoxide anion (O₂⁻) resulting in the production of NO₃ (Manahan, 1989 and Misko *et al.*, 1993). Various research studies (Jacob *et al.*, 1992; Elhassaneen, 2004; Elhassaneen *et al.*, 2020 and Elhassaneen *et al.*, 2023) have indicated that these substances are synthesized by the body during the progression of diseases such as arthritis, hepatotoxicity, obesity, anemia, cardiovascular ailments, and diabetes. An important discovery that lends support to the research hypothesis, suggesting a correlation between elevated oxidative stress-related disorders and CCl₄-induced liver damage, is the notable levels of MDA and NO₂ observed in this particular study. As such, the protection against hepatotoxicity might also be influenced by the antioxidative properties, lipid peroxidation, and inhibition of nitric oxide synthase by CTEE. This is evidenced by the considerable reduction in the production of MDA and NO₂ in the serum following dietary intervention. Detecting minimal concentrations of MDA and NO₂ would likely pose a challenge without any modifications in the antioxidant defense mechanism in rats administered with CTEE, in our assessment. The result agreed with Chayaratanasin *et al.*, (2019) and Maneesai *et al.*, (2021).

Table 6. Effect of intervention with CTEE on oxidative stress of the hepatotoxic rats induced by CCl₄

Value	Normal cont.	Model cont.	CTEE intervention (mg/kg bw/day)				
			25	50	100	200	400
Malondialdehyde concentration (MDA, nmol/mL)							
Mean	0.189 ^d	0.401 ^a	0.390 ^a	0.371 ^{ab}	0.335 ^b	0.266 ^c	0.220 ^e
SD	0.024	0.052	0.011	0.028	0.042	0.030	0.022
% of change	0.00	112.17	106.35	96.30	77.25	40.74	16.40
Nitrite (NO ₂ , nmol/L)							
Mean	2.32 ^d	4.95 ^a	4.85 ^a	4.36 ^b	3.59 ^c	3.10 ^b	2.82 ^e
SD	0.11	0.59	0.35	0.65	0.31	0.29	0.24
% of change	0.00	113.36	109.05	87.93	54.74	33.62	21.55

Each value represents mean \pm SD (n=6). Means on the same column with different superscript letters explain the significant difference at $P \leq 0.05$. Normal control, healthy rats without intervention; Model control, carbon tetrachloride-induced hepatotoxic rats without intervention; CTEE, *Clitoria ternatea* flower ethanolic extract; CTEE intervention, carbon tetrachloride-induced hepatotoxic rats with CTEE intervention.

Correlation studies

Significant discrepancies were observed in the correlation analysis between liver functions (AST, ALT, and ALP) and oxidative stress parameters (MDA and NO₂) and non-enzymatic antioxidant (GSH) in hepatotoxic rats induced by CCl₄ and feeding intervention CTEE Table 7. Based on the available data, a highly significant ($p \leq 0.05$) inverse association was observed between the serum concentrations of MDA and GSH ($r^2 = -0.947$). Furthermore, a substantial negative correlation ($p \leq 0.05$) was observed between liver functions (AST, $r^2 = +0.732$; ALT, $r^2 = +0.6955$; ALP, $r^2 = +0.663$ and TB, $r^2 = +0.814$) and MDA and NO₂ concentration in serum ($r^2 = +0.903$). The NO₂ evidence showed a similar pattern, though with lower values. A significant ($p \leq 0.05$) decrease in GSH, an antioxidant, and an increase in liver functions (AST, ALT, ALP, and TB) in rats serum as a result of hepatotoxic injury were found to be in opposition to an increase in oxidative stress parameters (MDA and NO₂). Argued alternatively, these correlations confirm to the issues of monitoring high concentrations of MDA and NO₂ in the absence of changes in the non-enzyme

antioxidant (GSH) of hepatotoxic rats. In accordance with Mehmood *et al.*, findings from (2019), an extract derived from the blue butterfly pea flower (*Clitoria ternatea* L.), that contains biologically active antioxidant aspects, impeded lipid peroxidation in hepatotoxic rats through elevated MDA and NO. According to Mehmood *et al.*, (2019) and Abd Elalal *et al.*, (2022), there was a relationship between relatively low concentrations of enzymatic and non-enzymatic antioxidants, such as GSH, and high levels of lipid peroxidation, particularly MDA and NO₂, in the plasma of hepatotoxic rats.

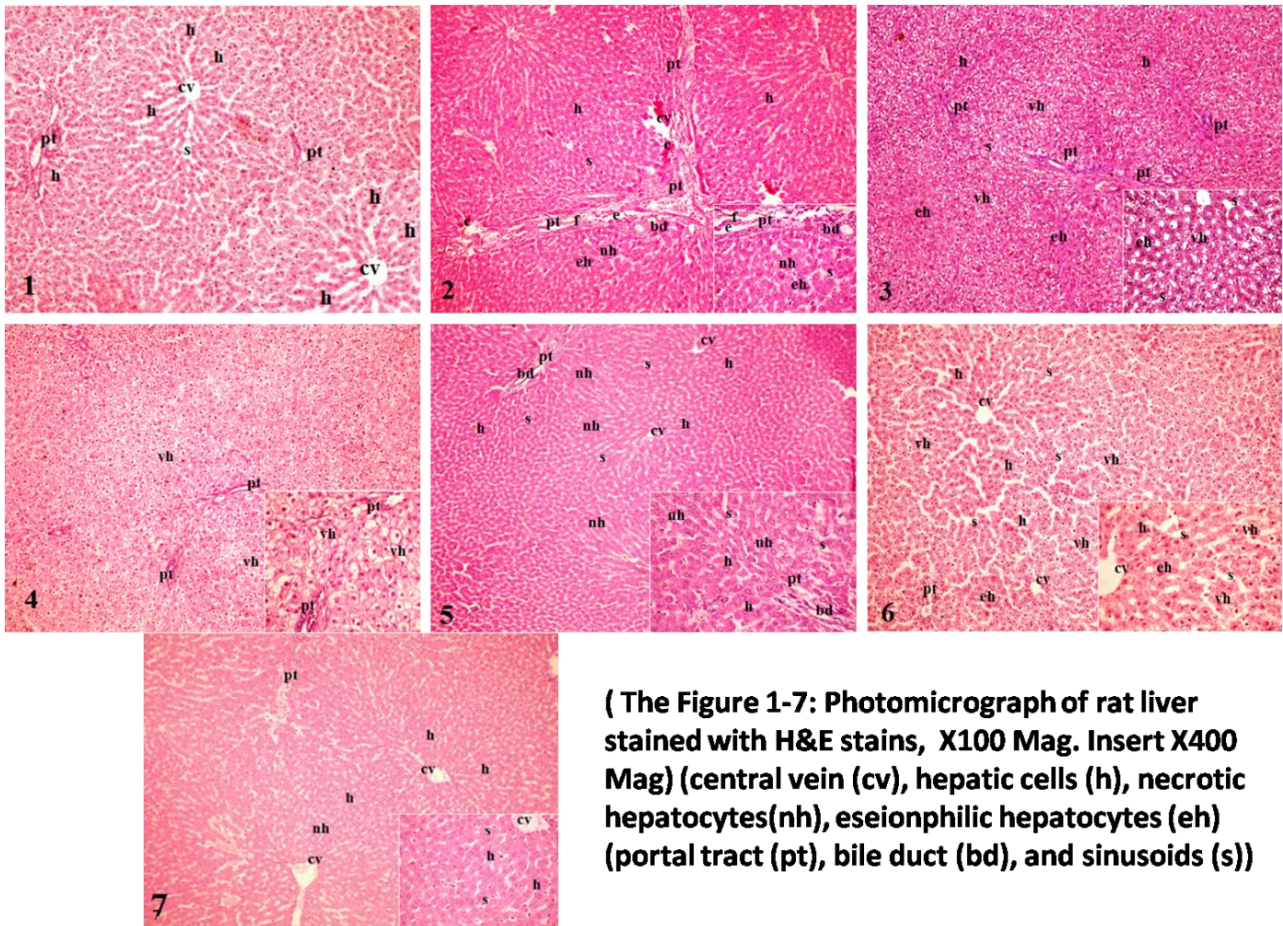
Histopathological Finding

Paraffin sections were stained with haematoxylin and eosin to study the histomorphological changes in rat live tissues in all groups. it was noticed the hepatocytes and stroma tissue have deep violet color of nuclei and pink cytoplasm. The white clear color may be indicted to the vacuolated cytoplasm or dissolving fats in the tissue section. The finding and discussion were illustrated as the following in Fig. 3(1-7):

Table 7. Correlation between oxidative stress and antioxidant parameters in hepatotoxic rats induced by CCl₄ and feeding intervention CTEE

Parameters	r^{2*}	Parameters	r^2
MDA/GSH	- 0.947	NO ₂ /GSH	- 0.891
MDA/NO ₂	+ 0.903	NO ₂ /AST	+0.693
MDA/AST	+0.732	NO ₂ /ALT	+ 0.597
MDA/ALT	+ 0.695	NO ₂ /ALP	+ 0.610
MDA/ALP	+ 0.663	NO ₂ /TB	+ 0.764
MDA/TB	+ 0.814		

* $P \leq 0.05$



(The Figure 1-7: Photomicrograph of rat liver stained with H&E stains, X100 Mag. Insert X400 Mag) (central vein (cv), hepatic cells (h), necrotic hepatocytes(nh), eosinophilic hepatocytes (eh) (portal tract (pt), bile duct (bd), and sinusoids (s))

1- normal control rat, show portal tract and central vein. Radiated hepatocytes from central vein with round nuclei and homogenous cytoplasm, mild dilation of sinusoids. 2- Model control rat liver administrated CCL_4 , note thick fibrotic area, three adjusting marked dilation congested portal tract with dilated bile duct attached with three congested central veins. There are infiltrating lymphocytes associated with many fibrotic cells and edema. The proliferating and crowded hepatocytes have round nuclei, other with necrotic nuclei (nh) and eosinophilic cytoplasm, mild dilation sinusoids was seen. 3- Rat liver administrated CCL_4 plus fed 25 mg of CTEE, note mild reorganized liver tissue, dilation and congestion portal tract with dilated bile duct, The hepatocytes (h) loss its attachments and proliferating through central vein. They have round nuclei with vacuolated cytoplasm. Many necrotic hepatocytes and dilation of sinusoids was seen. 4- Rat liver administrated CCL_4 plus fed 50 mg of CTEE, note mild reorganized liver tissue, reorganized portal tract and central vein. The proliferating hepatocytes through central vein and portal tract with small round nuclei and vacuolated cytoplasm and dilation sinusoids. 5- Rat liver administrated CCL_4 plus fed 100 mg of CTEE, note mild reorganized liver tissue, a mild dilation of portal tract with mild dilated bile duct. The hepatocytes was radiating and proliferating through central vein, they have round nuclei with homogenous cytoplasm and many necrotic hepatocytes, mild dilation of sinusoids was seen. 6- Rat liver administrated CCL_4 plus fed 200 mg CTEE, show mild dilation central vein. Radiated hepatocytes from central vein with round dark nuclei and homogenous cytoplasm, few eosinophilic hepatocytes and vacuolated cytoplasm hepatocytes, mild dilation of sinusoids. 7- Rat liver administrated CCL_4 plus fed 400 mg CTEE in die, show recovering and regenerative of central vein and portal tract. The hepatic cells have round dark nuclei and homogenous cytoplasm. They have organized radiating from central vein and portal tract with few necrotic hepatocytes sinusoids.

The experiment revealed that the CCL_4 facilitates the degenerative of hepatic tissues which resulted in (Figure 3.2) creation of massive fibrous septa, separation, and accumulation of pseudolobes of more than three portal vein attached to the central vein and formed a large fibrotic area. This finding was similarly (Sahreem *et al.*,2011 and Veidal *et al.*,2011) to the reported of many authors. In other hand the increased

of the CTEE doses in fed illustrated the recovering of the toxic effect of CCL_4 which appeared as recovering of most liver cell and the reduction of the fibrotic portal tract as well as the absent of infiltrating lymphocyte, fibrous cells and reduction of dilated portal tract and central vein and homogenous of most hepatocytes at group diet with 400 mg CTEE (Figure 3. 7). Therefore, the present results revealed that the CTEE

have ability for ameliorate the toxic effect of CCL₄ induced in liver tissue and its potential protective roll in tissue.

Conclusion

A beneficial protection against liver harm caused by CCl₄ was provided by *Clitoria ternatea* flower ethanol extract (CTEE). Our theory that various classes of bioactive compounds (phytochemicals) present in CTEE can break the hepatotoxic effects of CCl₄ by one or more of the following mechanisms was validated by these results: One approach to improve serum antioxidant defenses is by inhibiting excessive enzymatic activity that is expressed in liver functions. Another way is by reducing the amount of oxidative stress (MDA and NO₂) while boosting the formation of non-enzymatic antioxidants (GSH). There was the histopathological finding confirmed the biochemical result and revealed liver fibrosis induced by CCl₄ in rats and the recovering of the liver tissues with the high fed of the 200 - 400mg CTEE. That's why we recommended including extracts of this plant (*Clitoria ternatea* flowers) in our regular meals, beverages, and pharmaceutical formulae at concentrations of up to 400 mg/kg bw/day.

Abbreviations

ALP, alkaline phosphatase; ALT, alanine aminotransferase; AST, aspartate aminotransferase; BWG, body weight gain; CCl₄, carbon tetrachloride; CTEE, *Clitoria ternatea* flower ethanolic extract, FER, feed efficiency ratio; FI, feed intake; GSH, reduced glutathione; GSSG, oxidized glutathione; MDA, malondialdehyde; NO₂, Nitrite; TB, total bilirubin.

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REFERENCES

- Abd Elalal, N., S. Elsemelawy and Y. Elhassaneen. 2022. Potential Effects of Wild Milk Thistle (*Silybum marianum* L.) Seed Extract Intervention on Oxidative Stress Induced by Busulfan Drug in Different Organs of Rats. *International Journal of Healthcare and Medical Sciences*. 8(3): 19-34
- Abd El-Rahman, N. A. 2021. Potential effect of olive and mango leaves in diabetic rats induced by alloxane. MSc. Thesis in Nutrition and Food Science, Faculty of Home Economics, Minoufiya University, Shebin El-Kom, Egypt.
- Abushal, S.A. 2020. Hepatoprotective Effect of Moringa oleifera Leaves and Seeds Powder on Carbon Tetrachloride Induced Hepatotoxicity in Rats. *Alex. Sci. Exch. J.*, 41pp.371-379.
- AIN. American Institute of Nutrition. 1993. Purified diet for laboratory Rodent, Final report. *J. Nutrition*. 123:1939-1951.
- Almaadawy, A., R. Arafa and Y. Elhassaneen. 2016. Oxidative Stress and antioxidant defense systems status in obese rats feeding some selected food processing by-products applied in bread. *Journal of Home Economics*. 26 (1): 1 -37.
- AOAC 1990. Official Methods of Analysis of the Association of Official's Analytical Chemists, 14th Ed. Association of Official Analytical Chemist, Washington D.C., pp: 223-225, 992-995.
- Arya, M. R., Suhara Beevy S, Mathew Dan. 2022 .Pharmacological and Phytochemical studies on *Clitoria ternatea* L. –A review. *International Journal of Botany Studies*, 7: 446- 471
- Bera, T.K., K. Chatterjee, K. Jana, K.M. Ali, D. De and S. Maiti. 2012. Antihepatotoxic effect of “*L. ivshis*,” a polyherbal formulation against carbon tetrachloride-induced hepatotoxicity in male albino rat. *J Nat Pharm.*, 3: 17-24
- Boushey, C. J., A. M.Coulston, C. L.Rock and E. Monsen. (Eds.). 2001. *Nutrition in the Prevention and Treatment of Disease*. Elsevier.
- Buege, J.A. and S.D. Aust.1978. Microsomal lipid peroxidation in Packer L., (ed), *Methods in enzymology*, New York, NY, Academic, 52: 302 - 310.
- Chakraborty, G. S., V. Kumar, S. Gupta, A. Kumar, N., Gautam and L. Kumari. 2018. Phytochemical and pharmacological aspects of *Clitoria ternatea*-a review. *Journal of Applied Pharmaceutical Sciences and Research*, 3-9.
- Chapman, D.G., R. Castilla and J. A. Campbell. 1959. Evaluation of protein in food. I.A. Method for the determination of protein efficiency ratio. *Can. J. Biochemistry Physiology*, 37: 679-686.
- Chayaratanasin, P., A. Caobi, C.Suaprom, S. Saenset, P. Pasukamonset, N.Suanpairintr and S. Adisakwattana. 2019. *Clitoria ternatea* flower petal extract inhibits adipogenesis and lipid accumulation in 3T3-L1 preadipocytes by downregulating adipogenic gene expression. *Molecules*, 24(10), 1894.
- Cheeseman, K. H. and T.F. Salter. 1993. An introduction to free radical biochemistry. *Br Med Bull.*, 49(3):481-93.
- Cline, S., N.R.James, T. Silvia, J. Lawrence and C. Philip. 2004. Malondialdehyde adducts in DNA arrest transcription by T7 RNA polymerase and mammalian RNA polymerase II . *PNAS*, 101 (19): 7275–7280.
- Drury, R.A.B. and E.A. Wallington. 1980. *Carleton's histological technique*, fifth edition, Oxford University press, New York, Toronto, Hong Kong and Tokyo.36-125.
- Elhassaneen, Y. 1996. Chemical and technological studies on the pollution of environment with pesticides and polycyclic aromatic hydrocarbons. Ph.D. Thesis, Faculty of Agriculture, Mansoura University, Mansoura, Egypt.

- Elhassaneen, Y. A., S. A. Khader and M. A. El-aslowty. 2023. Potential Ameliorative Effects of Graviola (*Annona muricata* L.) Fruits on Carbon-tetrachloride Induced Hepatic Injury in Rats: Antioxidant, Apoptotic, Anti-inflammatory Markers, and Histopathological Studies. *International Journal of Healthcare and Medical Sciences*, 9(2): 17-31
- Elhassaneen, Y., S. El-Dashlouty and N. El-Gamal, 2020. Effects of brown algae (*Sargassum subrepandum*) consumption on obesity induced changes in oxidative stress and bone indices. *Journal of Home Economics*, 30 (4): 687-708.
- Elhassaneen, Y., A. Abd El-Rahman and S. El-Samouny. 2021-a. Potential Protective Effects of Cauliflower Leaves and Prickly Pear Fruits Skin on Liver Disorders Induced by Carbon Tetrachloride in Rats. *Journal of Home Economics*, 32 (1): 19-42.
- Elhassaneen, Y., M.S. El-Dashlouty, A. El-Eskafy and N. El-Gamal. 2016. The Potential Protection Effects of Cape Gooseberry (*Physalis peruviana* L.) Feeding Against CCl₄-Induced Hepatotoxicity in Rats. *Research Journal of Medicine and Medical Sciences*, 11(2): 1-13. [ISSN: 1816-272X ; <http://arnmsmb.com/RJMMS/>]
- Elhassaneen, Y., B.Hanaa, A. Abeer and B. Naglaa. 2021-b. Potential Effect of Milk Thistle on Liver Disorders Induced by Carbon Tetrachloride. *Journal of Home Economics*, 31 (1): 83-93.
- Elhassaneen, Y., Ragab, S. Thoraya, M. E. Azza and A. Abeer. 2012. Effect of Sweet Violet (*Viola odorata* L.) Blossoms Powder on Liver and Kidney Functions as well as Serum Lipid Peroxidation of Rats Treated with Carbon Tetrachloride. *The Journal of American Science*, 9 (5):88-95.
- Elhassaneen, Y.A. 2004. The effect of charcoal broiled meat consumption on antioxidant defence system of erythrocytes and antioxidant vitamins in plasma. *Nutrition Research*, 24 (6): 435 - 446.
- Ellman, G.L. 1959. Tissue sulphhydryl groups. *Archives of Biochemistry and Biophysics*, 82: 70-77
- ElSamouny, S. H. 2021. Potential protective effects of food industries by-products against liver disorders induced by carbon tetrachloride. Ph.D. Thesis in Nutrition and Food Science, Faculty of Home Economics, Minoufiya University, Shebin El-Kom, Egypt .
- Gella, F. J., T. Olivella, M. C. Pastor, J. Arenas, R. Moreno, R. Durban and J. A. Gomez. 1985. A simple procedure for the routine determination of aspartate aminotransferase and alanine aminotransferase with pyridoxal phosphate. *Clinica chimica acta*, 153(3):241-247.
- Grażyna, K., T. Piotr, P. Jan and S. Marta. 2020. Molecular Mechanisms of Ethanol Biotransformation: Enzymes of Oxidative and Nonoxidative Metabolic Pathways in Human. *Xenobiotica* 2020:1-56.
- Gupta G.K., J. Chahal and M. Bhatia. 2010. *Clitoria ternatea* (L) : Old and new aspects. *J Pharm Res* 3: 2610-2614.
- Hamzawy, M. A., E.S. El-Denshary, F.A. Mannaa and M.A. Abdel-Wahhab. 2013. Dietary supplementation of *Calendula officinalis* counteracts the oxidative stress and liver damage resulted from aflatoxin. *ISRN Nutrition*, 9(10):5402-538427.
- Jacob, T.D., M.K. Morrell, S. Manzi, J.B. Ochoa, V. Verdile, A.O. Udekwu, S.A. Berceli, R.L. Simmons and A.B. Peitzman. 1992. Nitric oxide: Implications for drug research. pp.28, IBC, South Natick, MA.
- Kebamo, S., T. Shibiru and G. Bekesho. 2015. The Role of Biotransformation in Drug Discovery and Development, *J Drug MetabToxicol.*, 5: 1-13.
- Khan, R. A., M. R. Khan and S. Sahreen. 2012. CCl₄-induced hepatotoxicity: protective effect of rutin on p53, CYP2E1 and the antioxidative status in rat. *BMC complementary and alternative medicine*, 12:1-6.
- Kuppan, N., Y. Kwan, Y. Lachimana, L. Subramanion, D. Ibrahim, C. Yeng, A. Chew and S. Sreenivasan. 2013., Evaluation of hepatoprotective effect of methanolic extract of *Clitoria ternatea* (Linn.) flower against acetaminophen-induced liver damage. *Asian Pac J Trop Dis.*, 3(4): 314-319.
- Lahon, K. and S. Das. 2011. Hepatoprotective activity of *Ocimum sanctum* alcoholic leaf extract against paracetamol-induced liver damage in albino rats. *Pharmacognosy Res.*, 3: 13-8
- Mahran, M.Z. and Y.E. Elhassaneen. 2023. Attenuation of Benzo[a]pyrene-Induced Oxidative Stress and Cell Apoptosis in Albino Rats by Wild Milk Thistle (*Silybum Marianum* L.) Seeds Extrac. *Egypt. J. Chem.* 66(SI: 13): 1671 - 1687.
- Manahan, S.E. 1989. *Toxicological chemistry: A guide to toxic substances in chemistry*, PCR press, New york.
- Maneesai, P., M. Iampanichakul, N. Chaihongsa, A. Poasakate, P. Potue, S.Rattanakanokchai and P. Pakdeecheote. 2021. Butterfly pea flower (*Clitoria ternatea* Linn.) extract ameliorates cardiovascular dysfunction and oxidative stress in nitric oxide-deficient hypertensive rats. *Antioxidants*, 10(4), 523.
- Mansour, Z. M. 2017. Hepatoprotective activity and antioxidant effects of avocado (*Persea americana*) fruits on rat's hepatotoxicity induced by carbon tetrachloride". M.Sc. Thesis in Nutrition and Food Science, Faculty of Home Economics, Minoufiya University, Egypt.
- Mee-Young, L., S. Chang-Seob, S. In-Shik, K. Young-Bum, K. Jung-Hoon. and S. Hyeun-Kyoo. 2013. Evaluation of oral subchronic toxicity of soshiho-tang water extract: the traditional herbal formula in rats. *Evidence-Based Complementary and Alternative Medicine*, . Article ID 590181, 9
- Mehmood, A., M. Ishaq, L. Zhao, S.Yaqoob, B. Safdar, M. Nadeem and C. Wang. 2019. Impact of ultrasound and conventional extraction techniques on bioactive compounds and biological activities of blue butterfly pea flower (*Clitoria ternatea* L.). *Ultrasonics sonochemistr.* 51:12-19.
- Meyer, S.A. and A.P. Kulkarni. 2001. Hepatotoxicity. In: *Introduction to Biochemical Toxicology*, Hodgson, E. and R.C. Smart (Eds.), 3rd Edn., John Wiley and Sons, New York, pp: 487-489

- Misko, T., R. Schilling, D.Salvemini, W. Moore and M. Currie.1993. A Fluorometric assay for the measurement of nitrite in biological samples. *Analytical Biochemistry*. 214: 11-16.
- Muhammad Ezzudin, R. and M. S.Rabeta. 2018. A potential of telang tree (*Clitoria ternatea*) in human health. *Food Research*, 2(5): 415-420.
- Mukherjee, P.K., V. Kumar, N.S. Kumar and M. Heinrich. 2008. The Ayurvedic medicine *Clitoria ternatea*-From traditional use to scientific assessment. *J Ethnopharmacol*. 120 (3): 291–301.
- Neda, G. D., M. S. Rabeta and M. T. Ong. 2013. Chemical composition and anti-proliferative properties of flowers of *Clitoria Ternatea*. *International Food Research Journal*, 20(3).
- Neeti J. N., C.C. Ohal, S.K. Shroff, R.H. Bhutada, R.S. Somani, V.S. Kasture, and S.B. Kasture, 2003. "Clitoria ternatea and the CNS". *Pharmacology Biochemistry and Behavior*. 75 (3): 529–536.
- Nilanjana M. 2013. Cardiovascular Diseases: Nutritional and Therapeutic Interventions.CRC Press., Taylor and Francis Group, London.
- Nithianantham K, K.Y. Ping, L. Latha, S.L. Jothy, I. Darah, Y. Chen, A.L. Chew. and S. Sasidharan. 2013. Evaluation of hepatoprotective effect of methanolic extract of *Clitoria ternatea* (Linn.) flower against acetaminophen-induced liver damage. *Asian Pac J Trop Dis.*, 3(4):314–9
- Nithianantham, K. , M. Shyamala, Y. Chen, L.YY. Latha, S.L.Jothy and S. Sasidharan, 2011. Hepatoprotective potential of *Clitoria ternatea* leaf extract against paracetamol induced damage in mice. *Molecules.*, 16(12): 10134-10145
- NRC, National Research Council. 1996. Guide for the Care and Use of Laboratory Animals Washington: National Academy Press.
- Pantazi, C. 2016. Watch this tea dramatically change from deep blue to vibrant red with a squeeze of lemon. *Business Insider Deutschland*. Archived from the original on September 30, 2018. Retrieved July 2, 2016.
- Parasuraman, S., R. Raveendran and R. Kesavan. 2010. Blood sample collection in small laboratory animals. *J Pharmacol Pharmacother*. 1(2):87-93.
- Parimaladevi, B., R. Boominathan, S. Mandal, C. Antiinflammatory. 2003.analgesic and anti-pyretic properties of *Clitoria ternatea* root, *Fitoterapia* 74: 345-349
- Patel, P. K. 2023. *Clitoria ternatea* L. var. *angustifolia* Hochst. ex Baker (Fabaceae): new varietal record to Asia from India. *Adansonia*, 45(14): 267-271.
- Pendbhaje, N.S., G. Sudheendra, S.M. Pathan, and D.S. Musmade. 2011. Thnopharmacology, pharmacogosity and phytochemical profile of *Clitorea ternatea* L inn: an overview. *Pharmacologyonline*, 3: 166-175.
- Pulok, M., V. Kumar, S. Kumar and N. Heinrich.2008. The Ayurvedic medicine *Clitoria ternatea*-From traditional use to scientific assessment. *Journal of Ethnopharmacology*. 120 (3): 291–301
- Rao, S. B., M. Jayanthi, R.Yogeetha, H. Ramakrishnaiah and J. Nataraj. 2013. Free radical scavenging activity and reducing power of *Gnidia glauca* (Fresen.) Gilg. *Journal of Applied Pharmaceutical Science*, 3(6): 203-207.
- Ross, A. C., B.Caballero, R. J.Cousins and K. L. Tucker. 2020. *Modern nutrition in health and disease*. Jones & Bartlett Learning.
- Sahreen, S., K. Muhammad and A. Khan. 2011. Hepatoprotective effects of methanol extract of *Carissa opaca* leaves on CCl₄-induced damage in rat. *BMC Complementary and Alternative Medicine*, 11:48-55
- Sarma, D. S. K., D. Kumar, C.Yamini, C. Santhalahari, C. Lahari, G. C. Kumar and M. Lahitha. 2023.Review on *Clitoria Ternata*. *Int. J. Pharm. Sci. Med*, 8(9): 43-58.
- Shanmugasundaran, R., V.K. Devi, P.S. Tresina, A. Maruthupandian, V.R. Mohan.2010. Hepatoprotective activity of ethanol extracts of *Clitoria ternatea* L. and *Cassia angustifolia* vahl leaf against CCl₄ induced liver toxicity in rats. *IJRP* 1(1):201–205
- Snedecor, G. W. and W.G. Cochran. 1967. *Statistical Methods"* 6th Ed. Iowa State University press. Ames. Iowa. USA.
- Solanki, Y.B. and S.M. Jain. 2012. Wound Healing Activity of *Clitoria ternatea* L. In *Experimental Animal Models*. *Pharmacologia*, 3: 160-168.
- Sreeharini, M., J. P. Yanadaiah, T. Lakshman and S. Durga Prasad. 2013. Hepatoprotective activity of aqueous ethanolic extract of roots of *Clitoria ternatea* Linn against carbon tetrachloride induced hepatotoxicity in rats. *J Pharm Biol Sci*, 1(4): 45-49.
- Susilo R, J.K., D. Winarni, S.A. Husen, S. Hayaza, H. Punnapayak, S.P. Wahyuningsih, E.S. Sajidah and W. Darmanto. 2019. Hepatoprotective effect of crude polysaccharides extracted from *Ganoderma lucidum* against carbon tetrachloride-induced liver injury in mice, *Veterinary World*, 12(12):1987-1991.
- Tahoon, S. R. 2019. Hepatoprotective effect of Apricot and Plum kernel on carbon tetrachloride induced hepatic rats ". *M.Sc. Thesis in Nutrition and Food Science*, Faculty of Home Economics, Minoufiya University, Egypt.
- Thilavech, T., S. Adisakwattana, P. Channuwong, K.Radarit, K.Jantarapat, K. Ngewlai and C. Chusak. 2021.Clitoria ternatea flower extract attenuates postprandial lipemia and increases plasma antioxidant status responses to a high-fat meal challenge in overweight and obese participants. *Biology*, 10(10):975.
- Tietz, N.W., E.L. Pruden and O. Siggaard-Anderson. 1994. In: *Tietz Textbook of Clinical Chemistry*. W.B Saunders Company London. Pp: 1354-1374
- Vassault, A., D. Grafmeyer, J.Graeve, R.Cohen, A. Beaudonnet, and J. Bienvenu. 1999. Quality specifications and allowable standards for validation of methods used in clinical biochemistry. *Ann Biol Clin (Paris)*., 57(6): 685-95.
- Veidal, S., M. Karsdal, E. Vassiliadis, A. Nawrocki, M. Larsen, Q. Nguyen, P. Häggglund, Y. Luo, Q. Zheng, B. Vainer and D. Leeming.2011. MMP mediated degradation of type VI collagen is highly associated with liver fibrosis

- identification and validation of a novel biochemical marker assay. *PLoS one*, 6 (9): 247-53.
- Vidana Gamage, G. C., Y. Y. Lim and W. S. Choo. 2021. Anthocyanins from *Clitoria ternatea* flower: Biosynthesis, extraction, stability, antioxidant activity, and applications. *Frontiers in Plant Science*, 12, 792303.
- Vuong, T. and H. Parichat. 2021. "Influences of pH on binding mechanisms of anthocyanins from butterfly pea flower (*Clitoria ternatea*) with whey powder and whey protein isolate". *Cogent Food & Agriculture*.7 (1): 1889098.
- Wang, L., X. Fang, B. Ling, F.Wang, Y. Xia, W.Zhang and X.Wang. 2024. Research progress on ferroptosis in the pathogenesis and treatment of neurodegenerative diseases. *Frontiers in Cellular Neuroscience*, 18, 1359453.
- Weber, L. W., M. Boll and A.Stampfl. 2003. Hepatotoxicity and mechanism of action of haloalkanes: carbon tetrachloride as a toxicological model. *Critical reviews in toxicology*, 33(2), 105-136.
- WHO. 1999. World Health Organization. "Environmental Health Criteria 208: carbon tetrachloride, Geneva, Switzerland,
- Yu, G., I.V. Nizhenkovska and M.M. Korda. 2020. Biological and Bioorganic Chemistry: in 2 books. Book 2. Biological Chemistry; edited by Yu.Gubsky, I.V.Nizhenkovska. Kyiv: AUS Medicine Publishing, 544 p.

الملخص العربي

التأثيرات الوقائية المحتملة للمستخلص الإيثانولي لزهرة البازلاء الفراشة (*Clitoria ternatea* Linn) ضد التسمم الكبدي الناجم عن رابع كلوريد الكربون في الفئران

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أجريت الدراسة الحالية لاستكشاف التأثيرات الوقائية المحتملة للمستخلص الإيثانولي لزهرة بازلاء الفراشة (*Clitoria ternatea* Linn) ضد التسمم الكبدي الناجم عن رابع كلوريد الكربون في الفئران. ولقد اوضحت النتائج انه عند المقارنة بالفئران الطبيعية ، أظهرت الفئران المحقونة برابع كلوريد الكربون (CCl₄) انخفاضا معنويا (p≤0.05) في المعايير البيولوجية المختلفة بما في ذلك الزيادة في وزن الجسم، المدخول من الغذاء، ونسبة كفاءة بمعدل -٤٢,٨٦%، و-٣٦,٩٩ و-٣٠,١٤% على التوالي. كما أظهرت المؤشرات الكيموحيوية مثل نشاط إنزيمات الكبد (ALT ، AST ، ALP) والبيلبيروبين الكلي ارتفاعاً معنوياً (p≤0.05) عند مقارنتها بفئران المجموعة الطبيعية بمعدل ٧٧,٢٨ ، ١٢٩,٩٠ ، ١٥٨,٧٢ و ٤١٢,٨٤% على التوالي. وقد تزامن ذلك أيضاً مع حدوث خلل في بعض مؤشرات الأوكسدة/مضادات الأوكسدة في الدم، أي انخفاض في مستوى مضادات الأوكسدة غير الأنزيمية (GSH و GSSG) ، وزيادة في مستوى المواد المؤكسدة (MDA ، NO₂) وكل هذه العوامل تشير إلى أن إصابة الكبد بمركب رابع كلوريد الكربون. كما أظهرت التدخل بالمستخلص الإيثانولي لزهرة البازلاء الفراشة في بروتوكول تغذية الفئران تحسينات معنوية (p≤0.05) كبيرة في جميع المقاييس البيولوجية والكيميائية الحيوية والمؤكسدات/مضادات الأوكسدة السابقة والتي تشير إلى الحماية ضد السمية الكبدية في الفئران. كما أن معدل التحسن في جميع هذه العوامل أظهر بطريقة تعتمد على الجرعة . وفي النهاية ، كان المستخلص الإيثانولي لزهرة البازلاء الفراشة فعالاً في الحماية من اضطرابات الكبد الناجمة عن رابع كلوريد الكربون ، ولذلك، توصى الدراسة بان يتم تضمين مثل هذه المستخلصات من أجزاء النبات (زهرة *Clitoria ternatea*) بتركيزات تصل إلى ٤٠٠ ملجم/كجم من وزن الجسم/اليوم في وجباتنا الغذائية والمشروبات والتركيبات الدوائية اليومية.

الكلمات المفتاحية: زهرة بازلاء الفراشة ، وزن الجسم، وظائف الكبد، أجزاء الجلوتاثيون، المالونالدهيد، النتريت.