

# Jojoba Leaf Extract Induced Nematocide Effect Upon *Meloidogyne incognita*

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## ABSTRACT

This work aims at disclosing the capability of Jojoba leaf extract to induce nematocide effect upon *Meloidogyne incognita*. To achieve such a purpose adventitious roots of *Allium cepa* were employed. Furadan as a well known nematocide was chosen and used for the comparison. Tomato seedlings were treated with *M. incognita* and soil was treated either with Furadan or Jojoba leaf extract. Mitotic index and chromosomal abnormalities were estimated. The obtained results revealed that the Jojoba leaf extract was proven to be active as nematocide agent.

## INTRODUCTION

In Egypt, jojoba is considered as one of the most practical solutions for desert plantation because its ability for tolerance to heat, drought and salt, lesser possibilities for infection, lesser need for fertilizers, and generous financial income, are certainly the most encouraging goals to plant jojoba in Egypt for revegetation of arid areas as it can survive in harsh desert environments; this is so in Mexico, Israel and in Rajasthan India, so that El Moguy (2002) was the first who mentioned the first practical steps to introduce jojoba to Egypt by planting the first jojoba field surrounded by date palms with the technical help of an agricultural expert, and the help of the FAO coordinator. The plant is important to commerce as its seeds oil store (40-60 % by dry weight). Its seed oil is a unique liquid wax ester mixture highly marketable for cosmetics and lubricants. It has promising physical properties, such as high viscosity index, high stability and freezing point, and can be used in various industries. It does not get damaged or rancid by repeated heating to temperatures above 300 °C.

The viscosity index of jojoba oil is much higher than that of petroleum oil, and it is therefore being used as a high temperature and high pressure lubricant in heavy machinery. In addition to this, it is also being used in transformer oil, detergents, the leather and plastic industries, and in pharmaceutical as well as cosmetic industries. Jojoba oil contains straight- chained C<sub>20</sub> and C<sub>22</sub> fatty acids and alcohols and two unsaturated bonds, which make the oil susceptible to many different types of chemical manipulations. The extracted oil is relatively pure, non-toxic, biodegradable, and resistant to rancidity.

On the other hand, the seed meal remaining after extraction of the oil has some uses it is mostly discarded. The rest of jojoba seed contains 25-30% crude protein and has potential as an animal feed supplement (Flo *et al.*, 1998; Jones and Lewis, 1999 & 2001; Brown, 2003 and Motawe, 2006) or partial replacement of fish meal by jojoba meal supplemented with Methionine and Biogen<sup>®</sup> for fish feeding (khalil *et al.*, 2009).

Jojoba is a promising oil crop and as the bushes are perennials, there will be a continuous supply of seed annually. This, and other advantages of the jojoba trees, such as soil conservation and as wind breaks add tremendously to the values of jojoba and to ensure further development of jojoba as a commercial crop. It is crucial to identify the factors that contribute to the extreme variability observed in different genotypes. Identification of genotypes capable of ensuring both profitable yield and wide genetic variability will be a challenging task that will require a complete set of information in order to understand how a given phenotype is constituted at the molecular. Biochemical, reproductive and agronomic level, thereby facilitating the rapid identification of molecular and metabolic markers that are important to define a required phenotype. A multidisciplinary approach based on molecular genetics, functional genomics, plant reproductive biology, biochemistry and agronomy will provide accurate information with which to identify genotypes with stabilized yield in various production systems. In vitro clonal propagation has a major advantage over seed propagation in that it allows propagation of unique genotypes which have been identified as being desirable. It allows propagating plants of known sex, or of known high-yield potential, or of other desirable characteristics.

This study was planned to investigate the effect of nematocide (Furadan 10G) on the nematode development compared with the effect of Jojoba leaves. However to disclose the capability of Furadan in inducing genotoxic effect on higher plant, Onion (*Allium cepa*) was selected, treated and two genotoxic bioassays were employed. They are : 1- investigation of cell proliferation, and 2- analysis of mitotic chromosomal behavior

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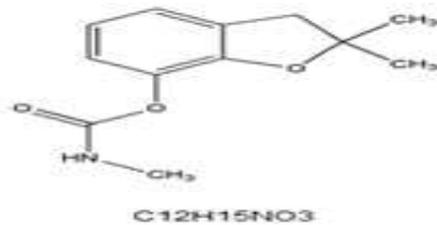
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Received May 9, 2013, Accepted June 26, 2013

## MATERIALS AND METHODS

### Furadan :

#### Structural Formula



**Chemical name:** (2,3-dihydro 2,2-dimethyl- 7-benzofuranyl methylcarbamate)

**Source :** (Kafr El-zayat) Company

#### Root-Knot nematode inoculum preparation:

Root-Knot nematode, *Meloidogyne incognita* used in the present study was originally isolated from galled tomato roots. Identification of the nematode to species was based on the morphological characteristics of the perineal patterns of adult females according to the key given by Taylor and Sasser (1978).

#### Effect of Jojoba dry leaves on the development and reproduction of *M.incognita*:

Effect of jojoba " *Simmondsia chinensis*" dry leaves on development and reproduction of *M. incognita* was studied. Twenty pots (10cm, diameter) were used in this experiment, each pot contains two tomato (cv. Super Strain B) plants. Pots were divided into five groups each of four replicates. The 1<sup>st</sup> and 2<sup>nd</sup> group received milled jojoba dry leaves at two concentrations 2.5 and 5g/ pot, respectively. Pots of the 3<sup>th</sup> and 4<sup>th</sup> groups received the granular nematicide Furadan® (10G) z-hich was applied in two concentrations 1.5 and 3g/ pot. The 5<sup>th</sup> group was left untreated to serve as a control. Treatments were applied in a hols around the tomato roots, 7 days after tomato transplantation.

After 48 hours from treatments by jojoba or Furadan®. Plants were inoculated by 5000 eggs/ pot in a hols around roots of plants.

The fertilizer, (N. P.K) (20,12,16%) at the rate of 2g/L was used as foliar application once every 10 days. Sixty days after nematode inoculation, tomato roots were gently removed from the soil and washed with tap water. Number of root galls, egg masses and 2<sup>nd</sup> stage juveniles/ 250g soil were recorded.

#### Slide preparation and staining:

The fixed root-tips were thoroughly washed with distilled water then .a single root- tips was placed in a drop of acetocarmine on a dry slide and covered.

### Studying the effect of root-knot nematode, *M.incoanita* on nuclear DNA content of the infected tomato roots:

Tomato seeds were allowed to germinate until an average of seedling height of 10 cm had reached. Then seedlings were transferred into plastic pots (10 cm diameter) each contained two seedlings.

The experiments were divided into three groups. The 1<sup>st</sup> and 2<sup>nd</sup> groups were inoculated with *M incognita* at the rate of 2000 J<sub>2</sub>/pot on the 3<sup>rd</sup> and 8<sup>th</sup> day after transplantation; the 3<sup>rd</sup> group was left noninoculated to serve as a control. Tomato seedlings were inoculated with *M incognita* 2000 J<sub>2</sub> /pot, Samples of cells were then taken after 15 days from tomato root- tips.

## RESULTS AND DISCUSSION

The present work aims to investigate the capability of *Meloidogyne incognita* in inducing differential DNA content in higher plant genome. The genotoxic effect of the nematicide Furadan® upon higher plant chromosomes was also investigated. A comparison between the effect of Furadan® and Jojoba leaf extract on the *Meloidogyne* development was carried out to investigate the nematocide effect of Jojoba leaf extract. In order to achieve such a purpose seedlings of tomato (*Lycopersicon esculentum*); and onion (*Allium cepa*) were chosen as higher plant test organism employing the following bioassays:

- 1- Flow cytometry for differential DNA content;
- 2- Cell proliferation estimated by measuring mitotic indices;
- 3- Analysis of mitotic chromosomal behavior; and
- 4- Investigation of *Meloidogyne* development.

The present result revealed that nematode populations were reduced with soil treatments of Furadan® (10G) a Concentration of 1.5 g and 3 g (a.i)/ pot, These data show that the nematicide Furadan® was proven to be highly effective on nematode reproduction. This conclusion is in accordance with that reported by Minton (1979), Inserra (1985), Siddqui *et al.*, (1998), Khan *et. al* (2001) and Amin (2013) .Comparing the effect of Jojoba leaf extract on the nematode development with Furadan®, The obtained results revealed that the Feuradan was effective significantly in reducing the number of galls, egg masses and 2<sup>o</sup>d stage juveniles of *M incognita* at two treatments 2.5 g and 5 g drv weight of leaves/ pot compared with the negative control group.

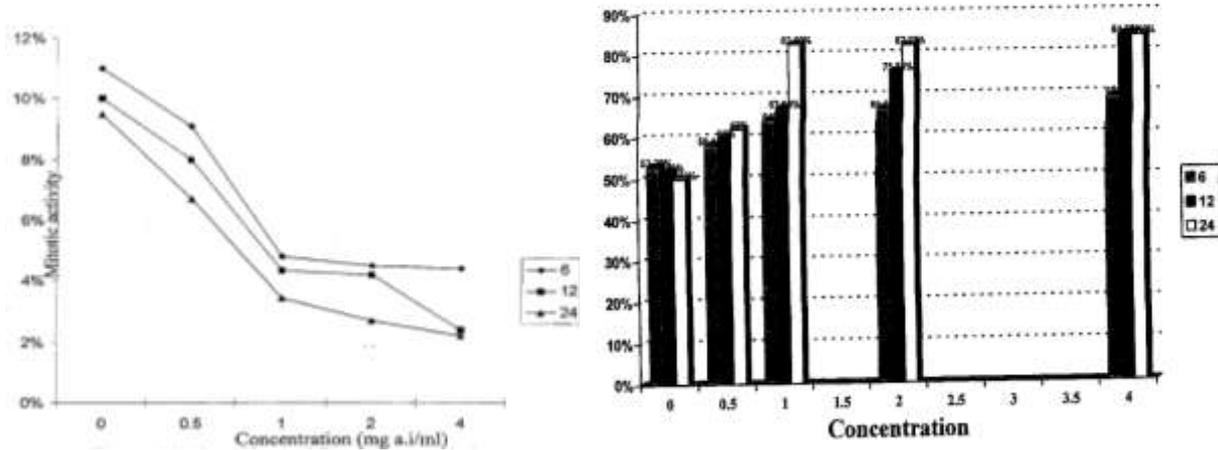
The present work suggests that the tested concentration of 2.5 g dry weight of Jojoba leaf extract was proven to be effective causing lethality for *Meliodyne incognita* more than that of Feuradan.

Regarding the genotoxic effect of Jojoba leaf extract upon *Allium cepa* genome, the present study showed that the tested Jojoba leaf extract has chromosomal damaging effect, since different types of aberrations were observed, these aberrations are deletion; gaps; breaks; fragments; stickiness; chromatide bridge; and binucleate. These types of aberrations however gave an evidence that Furadan® has a positive clastogenic activity upon the employed higher plant i.e., *Allium cepa* genome. Chromatide bridge gave an evidence that Jojoba leaf extract is capable to interfere and/ or alter the net change of the chromosomal proteins (Seehy and Hafez, 1994). This conclusion however is confirmed by the cytological observation of eroded surface obtained from this study. Binucleat cells presented an evidence that the tested nematocide is capable to prevent cell wall formation, and accordingly cytokinesis might be stopped.

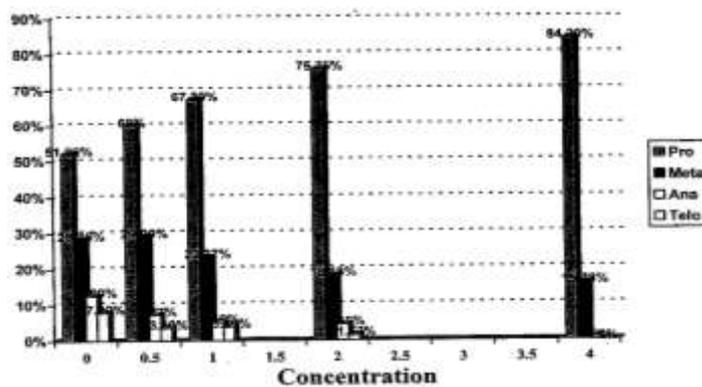
This result however presented an evidence that the tested Jojoba leaf extract has molecular activity upon chromosomes and cellular effect as well.

**Flow cytometry:**

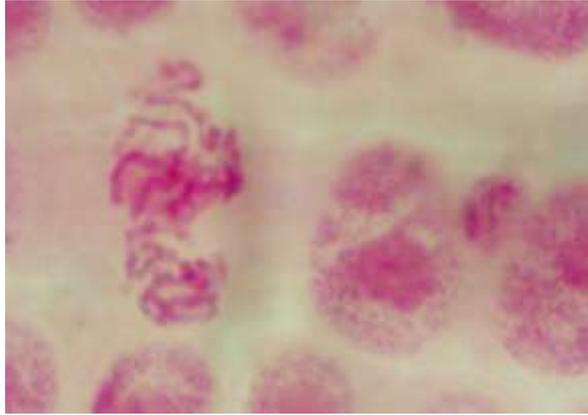
This step was achieved according to Kevers *et al.*, (1999) as following: suspension of isolated nuclei were kept cold on ice, and stain with an equivalent volume of 1 mg cm<sup>-3</sup> propidium iodide (PI) (Sigma, St. Louis, USA). This die was selected because it has an excitation spectrum (peak at 493 nm) compatible with the blue argonlaser present on most flow cytometers, and was found to be the best for determining DNA content in plant material (Michaelson *et al.*, 1991). Stained nuclei were analyzed directly in the dye solution. Nuclei were run on a FAC Star+ flow cytometer equipped with the FAC Station Computer Management System (Macintosh Cell Quest Software; Becton Dickinson, Sunyvale, USA) and with an argon- ion laser emitting a 488-nm beam at 200 mw of power (Spinnaker 1161 model; spectra- physics, mountain view, USA).



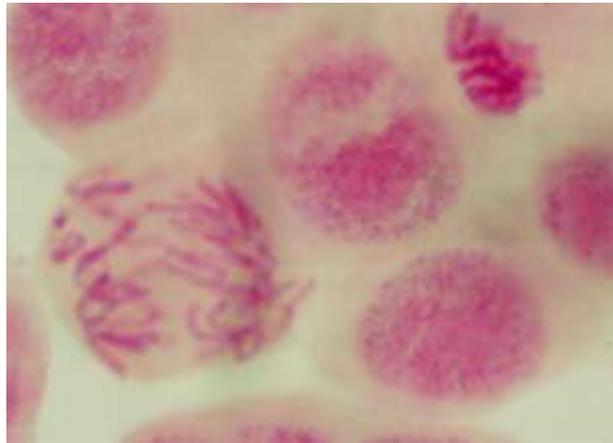
**Figure 1. The effect of Furadan( upon) the mitotic activity of *Allium cepa* root- tip cells after three treatment times (6, 12, and 24 hrs.)**



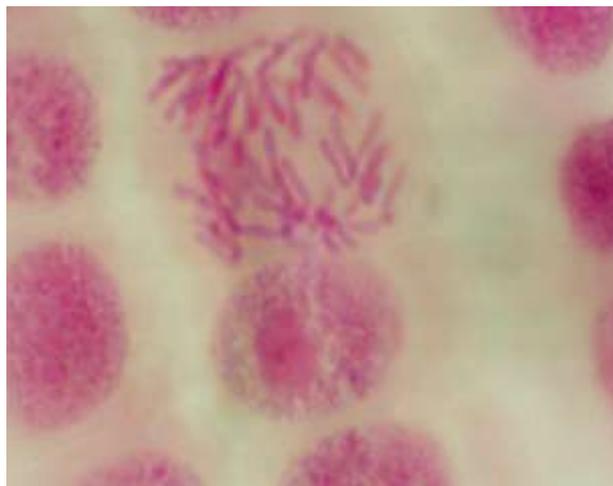
**Figure 2. The effect of 4 times concentration of Furadan on prophase indices for 3.**



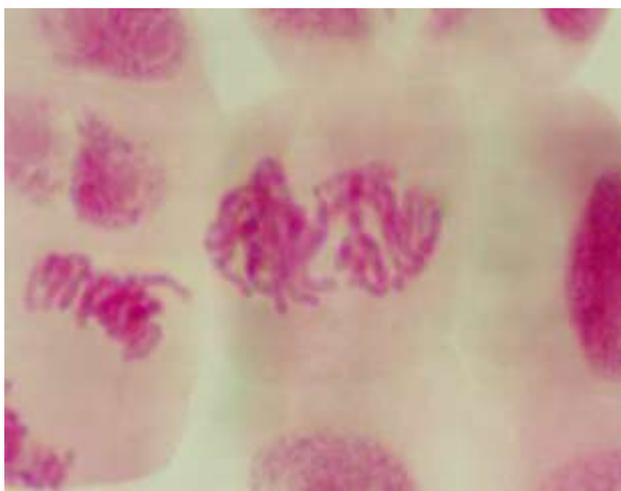
**Figure3.Phase indices of onion (*Allium cepa*) root- tip cells for 4 concentration at 12 treatment with Furadan.**



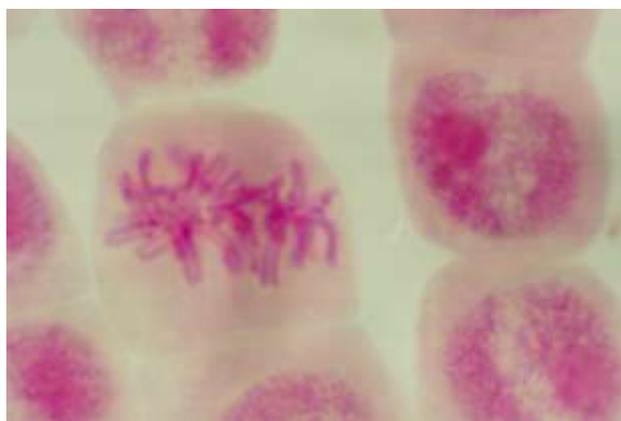
**Figure 4.Photomicrograph showing Anaphase with Stickiness and chromatid deletion after treatment with Furadan**



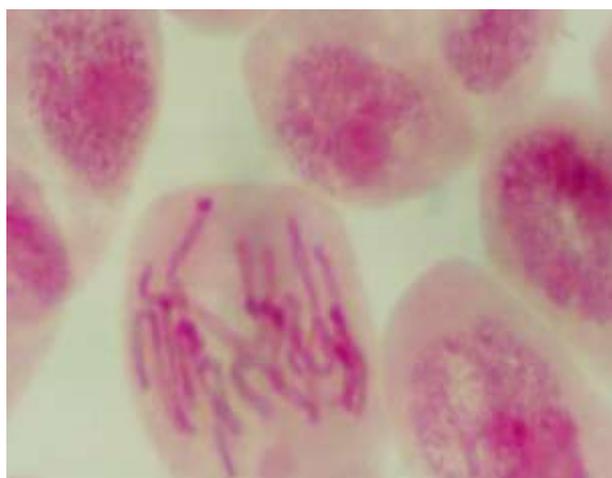
**Figure5.Photomicrograph showing Anaphase with chromatid deletion after treatment with Jojoba leaf extract**



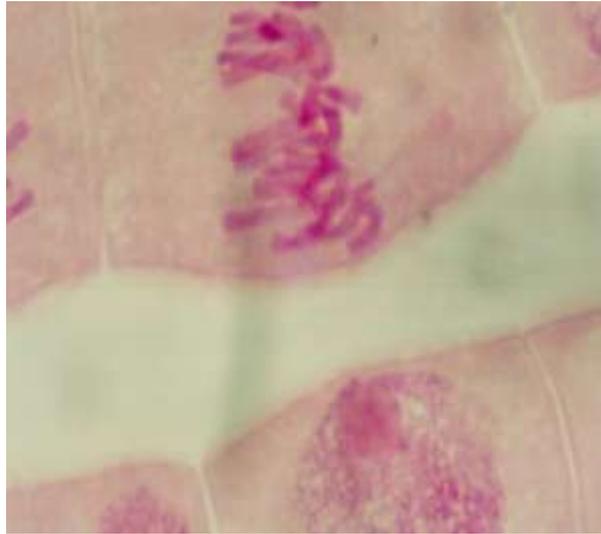
**Figure 6. Photomicrograph showing Anaphase with Stickiness chromatid deletion after treatment with Furadan**



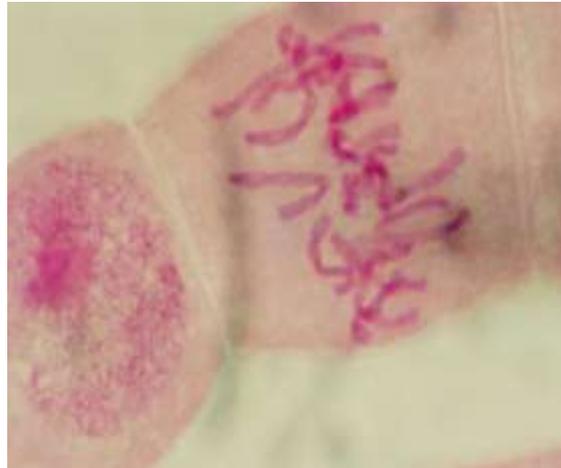
**Figure 7. Photomicrograph showing Tripolar Anaphase after treatment with Furadan**



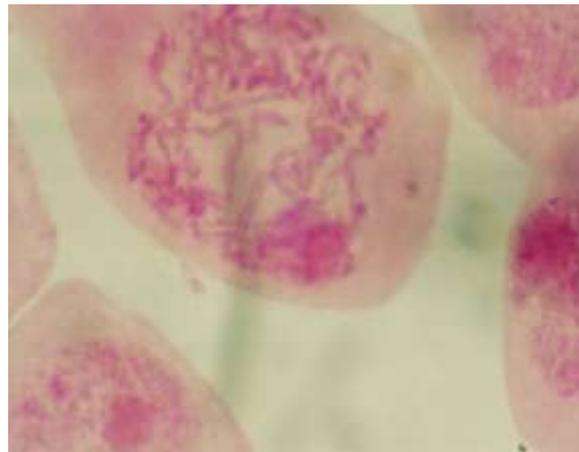
**Figure 8. Photomicrograph showing metaphase and gap**



**Figure 9. Photomicrograph showing metaphase with stickiness**



**Figure 10. Photomicrograph showing chromosome gap and chromosome fragment**



**Figure11. Photomicrograph of Prophase showing uncoiling**

## REFERENCES

- Abdel Maksud, S, A, (2003). Differential DNA Content and Development of Nematode. MS.c Thesis Fac. Of Agric. Alexandria University
- Amarger, V. and Mercier (1995). Molecular analysis of RAPD DNA based markers: Their potential use for the detection of genetic variability in Jojoba (*Simmondsia chinensis* L Schneider). *Biochimie* 77(12): 931-936.
- Amin, S. (2013). Genotoxicity of jojoba (*Simmondsia chinensis*) extracts employing a variety of short – term genotoxic bioassays. Ph.D. Thesis, Fac. Agric. Alexandria University.
- Bashir, M., M. Anjum and H. Rashid (2008). *In Vitro* propagation of some promising genotypes of jojoba (*Simmondsia chinensis*). *African Journal of Biotechnology* 7 (21): 3878-3886.
- Dolezel, J., Dolezelova, M., and Novak, F. J., (1994). Flow cytometric estimation of nuclear DNA amount in diploid bananas (*Musa acuminata* and *M. balbiana*) *Biologia Plantarum* 36 (3): 351-357.
- Insera, R. N., Vovlas, N., O'Bannon., J. H., and Griffin, G. D. (1985). Development of *Meloidogyne chitwoodi* on Wheat. *Journal of Nematology* 17 (3): 322 – 326.
- Kevers, C., Greimers, R., Franck, T., Bisbis, B., Dommes, J., and Gasper, T., (1999). Flow Cytometry estimation of nuclear of sugar beet. *Biological plantarum* 42 (3): 321 – 332.
- Khan, A., Shaukat, S. S. and Ahmed, I., (2001). Effect of organic Manures and Carbofuran on Nematodes Associated with Garlic ( *Allium Sativum* L.) Pakistan of *Biological Science*. 4 (3): 319 – 320.
- Michaelson, M. J., Price, H. J., Ellison, J. R., Johanston, J. S. (1991). Comparison of plant DNA contents determined by Feulgen microspectro photometry and laser flow cytometry. *Amer J. Bot.* 78: 183 – 188.
- Mills, D., Z. Yanqin and A. Benzioni (2009). Effect of substrate, medium composition, irradiance and ventilation on jojoba plantlets at the rooting stage of micropropagation. *Scientia Horticulturae* 121 (1): 113-118.
- Minton, N. A. and Admson, W. C., (1979). Control of *Meloidogyne Javanica* and *M. arenaria* on Kenaf and roselle with genetic resistance and nematocides. *Journal of nematology*, 11 (1): 37 – 43.
- Mohasseb, A. Heba, M. El-Bahr, Zakia, Adam, H. Moursy, and M. Solliman (2009). *In Vitro* Clonal Propagation of Jojoba (*Simmondsia chinensis* (Link) Schn.) *Australian Journal of Basic and Applied Sciences* 3(4):3128-3136.
- Sediqui, M.A., Azam, F. and Saxena, S.K. (1998). Evaluation of chemical Pesticides against the root-knot Nematode on Aubergine. *Ann. Appl. Biol.* 132.
- Seehy, A.M. and Hafez, M. A. (1994). Genotoxicity of contraceptive. 1st international conference on environmental mutagenesis. On human population at risk Egypt, Cairo, January, 1992, 19 – 24.
- Seehy, M., Y. A. Mostafa and M. A. Osman (1989). Evaluation of nystatin genotoxicity- Mitotic non-disjunction in *S. cerevisiae* and *Allium cepa* chromosomes. *Bull. Of Alex. Fac. Med.* Vol. Xxr no. 6: 1625-1631.
- Sharma, K., V. Agrawal, S. Gupta, R. Kumar and M. Prasad (2008). ISSR marker-assisted selection of male and female plants in a promising dioecious crop: jojoba (*Simmondsia chinensis*). *Plant Biotechnol Rep.* 2:239-243.
- Taylor, A. L., and J. N. Sasser (ed) (1978). *Biology identification and control of root-knot nematodes* North Carolina State University Graphic, Raleigh, NC, U.S.A 111 pp.

## الملخص العربي

### مستخلص أوراق الجوجوبا يحدث أثراً مضاداً للنيماتودا

أميرة خطاب ومحمد الصيحي

المادة الجافة/ أصيص يحتوي نباتين طماطم ثم عدوتها مسبقاً بـ 5000 بيضة من النيماتودا *M. incognita* وبعد مرور 30 يوم من المعاملة أوضحت النتائج قدرة ذلك المستخلص على خفض وتقليل عدد البيض وكذلك عدد كتل البيض وعدد اليرقات في كلا المعاملتين. ووجد أن الفيوردان كان أكثر قدرة على تقليل عدد البيض وعدد يرقات الطور الثاني وأيضاً عدد كتل البيض المتكونة على الجذور وفي حين كان تأثير الجوجوبا على النيماتودا بالتربة متساوياً تقريباً في المعاملتين إلا أنه أقل فاعلية من المبيد المستخدم.

ومن هذا يتضح أن مستخلص أوراق نبات الجوجوبا له أثر مضاد للنيماتودا وهذا يبشر بمستقبل جيد لهذا النبات ومستخلصاته.

هدف هذا البحث لدراسة أثر مستخلص نبات الجوجوبا كمضاد للنيماتودا وتم اختيار نبات الطماطم وتوظيف القياسات الضوئية المتدفقة علاوة على فحص الكروموسومات في القمم النامية للجذور العرضية لنبات البصل. وتم استخدام المبيد النيماتودي المعروف باسم الفيوردان للمقارنة. وتم توظيف المادة الوراثية لنبات البصل حيث أنه كشاف حساس لدراسة تأثير السمية الوراثية.

وكذلك اهتم البحث بمقارنة تأثير مستخلص أوراق نبات الجوجوبا على تطور النيماتودا وهو نبات منزرع في مصر حديثاً ومقارنة ذلك بتأثير الفيوردان على تطور النيماتودا أيضاً. وقد أوضحت النتائج أن استخدام مستخلص أوراق الجوجوبا بمعدل 2,5 جم، 5 جم من