

Effect of Certain Plant Extracts on Mortality, Development and Haemogram of *Culex pipiens* L. Mosquitoes Larvae

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ABSTRACT

The effect of methanolic plant extracts from *Solanum nigrum*, *Acokanthera spectabilis* and *Heliotropium aegyptiacum* were investigated on the larval mortality and development of *Culex pipiens* L. Also, the effects of the LC₅₀ of the three plant extracts on the larval haemogram were studied. Plant extracts exhibited variable bioactivities. The greatest activity was observed for *S. nigrum* which showed LC₅₀ values of 130.8 ppm after 48 hr of exposure, respectively. Percentage of larval mortality was 67.5% in the treatment with extracts of *S. nigrum* and 66.5% in case of *A. spectabilis* at 500ppm. Egg hatchability was not significantly reduced in all *S. nigrum* concentrations. All concentrations of the plant extracts from *S. nigrum*, *A. spectabilis* and *H. aegyptiacum* caused significant hindrance to the subsequent larval development and reduced both pupation and adult emergence. Drastic retardation of development was shown by *S. nigrum* extracts, where only 18.1% and 8.5% of the larval managed to reach pupal and adult stages, respectively, when reared in 100 ppm of the extract. However, *A. spectabilis* and *H. aegyptiacum* were more effective at higher concentrations. Moreover, the effects of the LC₅₀ of tested plant extracts caused a reduction in the number of haemocytes and also markedly decreased the haemocytes surface areas. Application of such plant extracts to mosquito breeding site may have great practical importance in relation to non-synthetic chemical control of this serious disease vectors.

INTRODUCTION

The house mosquito, *Culex pipiens* L. (Diptera: Culicidae) is one of the most harmful insects affecting humans and farm animals and transferring several pathogens as Cache-Valley (CV) and West Nile Virus that cause infertility and congenital malformations in ruminants (Edwards *et al.*, 1998 and Smartt and Erickson, 2008). This insect showed great resistance to the used synthetic insecticides by many mechanisms (Raymond *et al.*, 2001, Dary *et al.*, 1990). It also appeared to be sensitive to several plant extracts and products. Myrrh (oleo-gum-resin) obtained from the stem of *Commiphora molmol* proved to have insecticidal activity affecting fat, muscles, gut and nervous tissues of the larvae (Massoud and Labib, 2000). Methanolic extracts of aerial parts of the medicinal plant Argel, *Solenostemma argel* (Del.) Hayne, incorporated into rearing media of *Culex pipiens*

L. reduced oviposition, hatchability and larval viability (Al-Doghairi *et al.*, 2004). Cetin *et al.*, (2006) reported high larvicidal activity of *Teucrium divaricatum* Sieber, *Mentha longifolia* (L.) Huds., *M. pulegium* L., *Melissa officinalis* L. and *Salvia sclarea* L. oils against *C. pipiens*. Zhu *et al.* (2006) reported that the oils of thyme, catnip, amyris, eucalyptus and cinnamon revealed larvicidal activity besides repellent effect on this pest. Several plant species extracts killed and altered developmental periods, pupation rates and adult emergences of this insect (Khater and Shalaby, 2008). Several pure active potential sources of larvicidal substances against mosquito were isolated as beta-thujaplicin from *Chamaecyparis obtuse* leaves (Jang *et al.*, 2005), 7-hydroxycoumarin from *Stellera chamaejasme* root powder (Xiaorong and Taiping, 2008) and pure synthetic natural naphthoquinones (alkannin, shikonin and shikalkin) and three acetylated derivatives of shikonin (Michaelakis *et al.*, 2009). Control of *C. pipiens* is becoming increasingly difficult because the over production of detoxifying mechanisms of chemicals insecticides by this insect (Severini, *et al.*, 1993). The botanical insecticides are generally pest specific and are relatively harmless to non-target organisms including humans. They are also biodegradable and harmless to environment. The phytochemicals derived plant resources can act as larvicides, insect growth regulators, repellent and ovipositional attractant (Das *et al.*, 2003, Venkatachalam and Jebanesan, 2001). One plant species may possess substances with a wide range of activities, like extracts from *Azadiracta indica* which showed antifeedent, antioviposition, repellent and growth-regulating activity (Schmutterer, 1995). Insecticidal activity of many plants against several insect pests has been demonstrated (Carlini and Grossi-de-sa, 2002, Kundu, *et al.*, 2007 and Boussada, *et al.*, 2008).

In this study the effects of methanolic extracts of aerial parts from three plant species *Solanum nigrum*, *Acokanthera spectabilis* and *Heliotropium aegyptiacum* on egg hatchability, larval development and different haemocyte counts of *C. pipiens* mosquitoes, have been investigated.

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MATERIALS AND METHODS

Insects: A *Culex pipiens* L. (Diptera: Culicidae) colony maintained in the laboratory for more than 10 years was used. Mosquitoes were held at $27\pm1^{\circ}\text{C}$, $70\pm5\%$ RH and a photo period of 14:10 (light:dark). Adults were provided with a 10% sucrose solution as food source. A pigeon was introduced twice a week to the adults for blood feeding. Larvae were reared in dechlorinated water under the same temperature and light conditions and were fed daily with baby fish food. The experiment was carried out at the faculty of agriculture, Alexandria University, Egypt.

Plant extracts: Fresh fruits (1100 gm) of *Solanum nigrum*, *Acokanthera spectabilis* and *Heliotropium aegyptiacum* for each were collected from different places (Abbis, Wady el-Natrun) in Egypt. The collected samples were washed from dust and freshly blended with acetone (1.0 L) times for a month in the dark at room temperature. Fruits were air-dried for 5 days, ground to fine parts and extracted with 500ml of methanol at ambient temperature. A gentle warming to $35\text{--}40^{\circ}\text{C}$ was sometimes found necessary. The mixture was stirred for 30 minutes by magnetic stirrer and left 24 hr. Then, it was concentrated in a rotary evaporator under vacuum of solvent in a water bath at 55°C according to Chitra *et al.*, 199). The resulted filtrate was further concentrated to 0.25 L for each plant fruit.

Test procedure: Stock solutions of three plant methanolic extracts were prepared by dissolving the extracted in warm distilled water (at $0.5\text{g} / 100\text{ml}$ water) containing 0.5% Triton X100 as an emulsifier to ensure complete solubility of the extract in water. Different concentrations of 100, 200, 300, 400 and 500ppm were prepared from stock solutions. Ten 2nd instar larvae were transferred from the culture into plastic cups (8 cm diameter, 10 cm depth) each containing 30 ml of desired concentration. Treatments were triplicate and control had only distilled water. Larvae were fed daily and kept under laboratory conditions. 500 eggs were treated in water by the same previous concentrations of the plant extracts, 5 replicates (100 eggs for each). Egg hatchability was determined at 2 days after treatment. Larval mortalities were counted at 7 days after treatment. Percentage of successful pupation and adult emergence were determined by monitoring on daily basis until all adults in the control have emerged.

Haemocyte studies: The hemolymph of treated larvae with the LC_{50} of each tested plant extract were separately smeared to thin film between two glass slides. The blood smears were air-dried, stained with Wright's blood stain (Essawy, 1990) for 1 min and destained for 2 min. with 70% ethyl alcohol.

The larval hemocytes were examined using light microscope. Blood samples were obtained from 5 larvae of each extract and control. Each blood sample was replicated three times. The haemocyte types were examined and identified under oil immersion (100X) using a stereomicroscope. Different Haemocyte Count (DHC) was carried out in random scans of blood films (100 haemocytes from each film). The identification of haemocyte types were performed according to (Arnold and Hinks, 1983). In addition, the surface areas of each haemocyte type were measured by a micrometric slide in all treatments plus the control.

Statistical analysis: Data were analyzed using maximum likelihood procedures and values of LC_{50} were calculated according to finney (1971). Data were corrected for control mortality (Abbott, 1925). Data of egg hatchability were analyzed by analysis of variance. If significant differences ($p<0.05$) occurred, means were separated by Duncan's multiple range test.

RESULTS AND DISCUSSION

The mortality of *C. pipiens* larvae treated with three plant methanolic extracts and their LC_{50} values and 95% confidence limits at 48 hr after treatment are shown in Table 1. Results showed that the most toxic extract was *S. nigrum* followed by *A. spectabilis* and *H. aegyptiacum* at the concentration of 500 ppm, the larval mortality was 77.5% after 48hr of exposure in the case of *S. nigrum*, while it was 64.7% after 48 hr in the case of *A. spectabilis*, whereas the larval mortality was 21.7% after 48 hr in the case of *H. aegyptiacum*. LC_{50} s and 95% confidence limits (CL) for each plant are given in Table 1. Data showed a significant differences. LC_{50} for 2nd instar larvae were 130.8, 403.5 and 470.9 for *S. nigrum*, *A. spectabilis* and *H. aegyptiacum*, respectively, after 48 hr of exposure. *S. nigrum* was significantly more toxic at all exposure times than *A. spectabilis* and *H. aegyptiacum*.

Egg hatchability was not significantly reduced ($p<0.05$) by treatment with all extracts compared with control (Table 2). Whereas the larval mortality was significantly reduced. *S. nigrum* extract at 500 ppm revealed the most severe effect on larval mortality which compared with *A. spectabilis* and *H. aegyptiacum*. At concentration of 500 ppm, *S. nigrum*, *A. spectabilis* and *H. aegyptiacum* caused reduction in larval mortality of 67.5, 66.5 and 63.6%, respectively. The effect of the three plant extracts on growth and development of *C. pipiens* larvae to adulthood are given in Table 2. There were considerable reduction in the percentage of larvae undergoing successful pupation in all treatments compared with control. No further larval development took place after the 2nd instar in *S. nigrum* at 500 ppm.

Table 1. Mortality percentage of 2nd larvae of *Culex pipiens* larvae in media containing methanolic plant extracts at different exposure periods

Plant extract	Conc (ppm)	Mortality% after 48hr of exposure	Slope±S.E.	LC ₅₀ (95%CL)*
<i>Solanum nigrum</i>	100	46.1	1.2±0.8	130.8(95.8-190.1)
	200	54.6		
	300	57.2		
	400	65.9		
	500	77.5		
<i>Acokanthera spectabilis</i>	100	24.9	1.3±1.1	403.5(360-442)
	200	30.1		
	300	35.3		
	400	52.9		
	500	64.7		
<i>Heliotropium aegyptiacum</i>	100	10.1	1.5±2.3	470.9
	200	12.2		
	300	14.9		
	400	16.5		
	500	21.7		
Control	0	5.8	-	-

*LC₅₀ values and 95% confidence limits of 2nd instar larvae of *C. pipiens* larvae

Table 2. Egg hatchability, Larval mortality, Successful pupation and adult emergence of *Culex pipiens* larvae reared in media containing methanolic plant extracts

Plant extract	Conc (ppm)	Egg hatchability at 2 days after treatment	Larval mortality (%) at 7 days after treatment	Pupation (%)	Adult emergence (%)
<i>Solanum nigrum</i>	100	97.8 ^a ±1.8	16.6 ^b ±2.3	18.1 ^b ±1.2	8.5 ^b ±0.9
	200	97.6 ^a ±2.1	28.2 ^c ±1.9	15.3 ^b ±0.9	6.1 ^b ±1.5
	300	96.9 ^a ±1.9	46.9 ^d ±0.8	6.8 ^c ±1.5	4.2 ^c ±2.1
	400	96.7 ^a ±1.6	55.9 ^d ±2.1	5.3 ^c ±2.2	1.8 ^d ±1.4
	500	97.2 ^a ±3.1	67.5 ^e ±1.8	0.0 ^e	0.0 ^e
<i>Acokanthera spectabilis</i>	100	98.2 ^a ±2.1	13.2 ^b ±1.3	37.9 ^b ±0.8	12.8 ^b ±1.1
	200	97.9 ^a ±1.3	25.7 ^c ±1.6	20.1 ^c ±2.8	7.5 ^c ±2.2
	300	97.6 ^a ±1.6	42.9 ^d ±1.1	13.9 ^c ±1.7	4.9 ^c ±2.9
	400	97.5 ^a ±2.1	54.1 ^d ±1.7	7.2 ^d ±2.1	1.5 ^d ±1.2
	500	96.9 ^a ±0.9	66.5 ^e ±2.0	0.0 ^e	0.0 ^e
<i>Heliotropium aegyptiacum</i>	100	97.2 ^a ±1.7	11.8 ^b ±0.7	64.9 ^b ±1.1	36.9 ^b ±2.6
	200	97.5 ^a ±2.8	23.4 ^c ±1.3	55.9 ^b ±2.3	17.9 ^c ±3.1
	300	98.6 ^a ±3.1	39.8 ^d ±3.0	56.9 ^b ±0.9	16.9 ^c ±1.1
	400	96.8 ^a ±2.4	52.6 ^d ±1.1	29.5 ^c ±1.2	8.1 ^d ±0.8
	500	97.2 ^a ±1.3	63.6 ^e ±1.9	9.8 ^d ±2.2	3.2 ^e ±2.9
98.2 ^a	100.0 ^a	4.1 ^a	98.0 ^a	0	Control

Means followed by the same letter are not significantly different at 5% level, Duncan multiple test.

On the other hand all plant extracts had an evident inhibitory effect even at 100 ppm, whereas the successful pupation were only 18.1, 37.9 and 64.9% for *S. nigrum*, *A. spectabilis* and *H. aegyptiacum*, respectively. Complete suppression for adult emergence

was evident in case of *S. nigrum*, *A. spectabilis* at 500 ppm. *S. nigrum*, *A. spectabilis* and *H. aegyptiacum* at 100 ppm caused 8.5, 12.8 and 36.9% adult emergence, respectively.

Considerable biological activity related to toxicity and hindrance of growth and development of the larvae of *C. pipiens* has been observed in this study. *S. nigrum* was found to cause higher rate of mortality compared to other plant extracts. Srinivasan *et al.* (2013) found that *Solanum sarrachoides* was effective against green peach aphid. *A. spectabilis* and *H. aegyptiacum* exhibited a relatively mild acute effect on mosquito larvae especially in its lower concentrations. However, after 8 days of exposure the toxicity was almost high above 200 ppm. The larvicidal activity of some plant extracts, essential oils and phytochemicals against *C. pipiens* have been demonstrated (Traboulsi *et al.*, 2005; Abdelgaleil 2006; Michaelakis *et al.*, 2007; Radwan *et al.*, 2008).

Figure 1 showed the effect of the LC₅₀ of the tested plant extracts on the mean number of haemocytes in the haemolymph of 2nd instar larvae of *C. pipiens*. Four haemocyte types identified as Prohaemocytes (Pr), Plasmotocytes (Pl), Granulocytes (Gr) and Oenocytoides were monitored. With regard to *S. nigrum*, *A. spectabilis* and *H. aegyptiacum* treatments, a reduction in the percentage of the prohaemocyte was 74.6, 62 and 56.6, in respect less than control. The number of (Gr) was markedly decreased especially after treatment with *S. nigrum* and *A. spectabilis* with reduction percentages

of 48.8 and 43.6, respectively. The same trend was observed in the number of plasmotocyte which were reduced by 55 and 34.5 %, in respect. An increase was observed in the number of (Oe) about 45.16 and 61.3 %, in respect over the control. Furthermore, treatment with the LC₅₀ of tested plant extracts caused a decrease in the haemocyte surface areas.

The obtained results in (Fig., 2) proved that after the treatment with the LC₅₀ of *S. nigrum*, *A. spectabilis* and *H. aegyptiacum* methanolic extracts, the haemocyte surface areas were markedly decreased when compared to the control. The results indicated that treatments with *H. aegyptiacum* extract (Fig.3b) caused a decrease in all haemocytes surface area by about 33% in prohaemocyte, 11% in granulocyte, 13% in plasmotocyte and 7% in oenocytoides less than control. The same trend was observed in *A. spectabilis* extract treatment (Fig.3c) by about 42% in prohaemocytes, 21% in granulocyte, 20% in plasmotocytes and 19% in oenocytoides less than control. Also, treatment with *S. nigrum* extract caused a marked decrease in the haemocytes surface areas about 48% in prohaemocyte, 29% in granulocyte, 27% in plasmotocyte and 21 % in oenocytoides (Fig.3d) less than control. (Fig.3a)

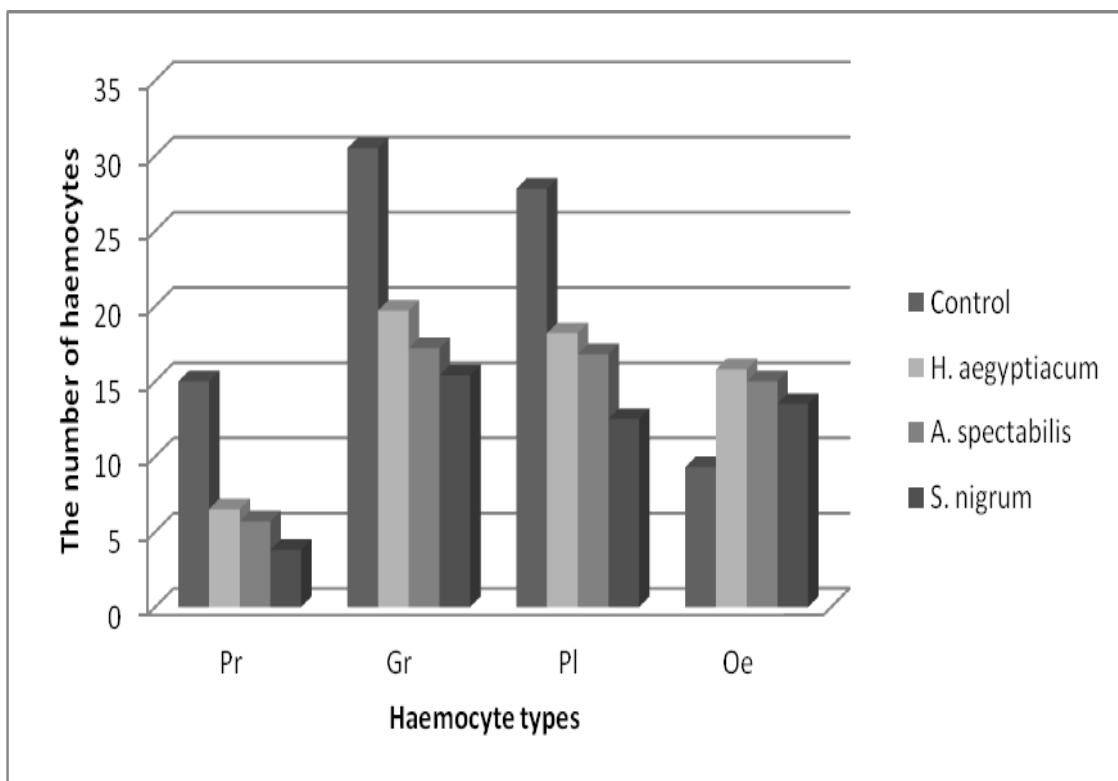


Figure1. Effects of methanolic plant extracts from *S. nigrum*, *A. spectabilis* and *H. aegyptiacum* on the differential haemocyte count in *C. pipiens* larvae

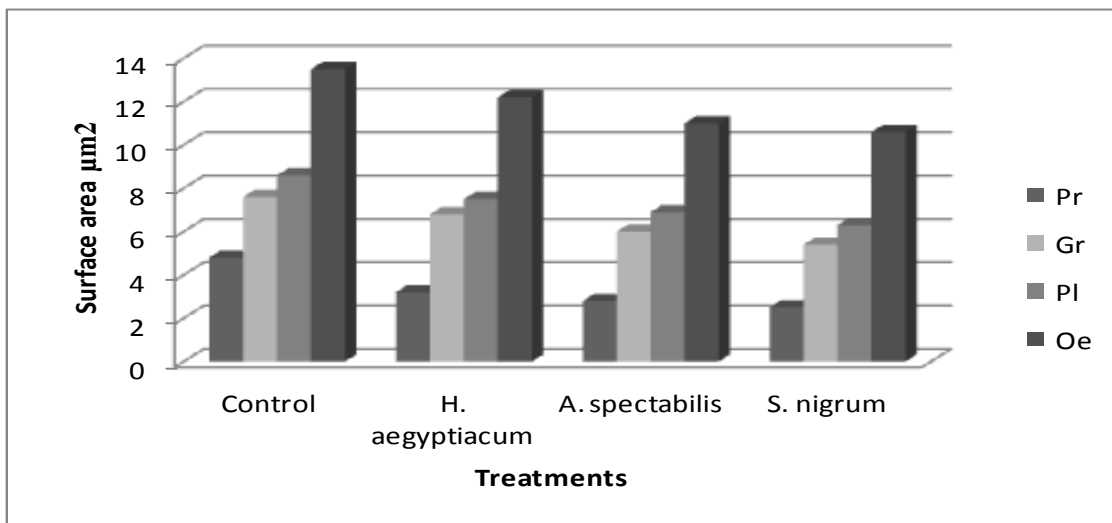


Figure2. Effects of methanolic plant extracts from *S. nigrum*, *A. spectabilis* and *H. aegyptiacum* on the haemocyte surface area in *C. pipiens* larvae

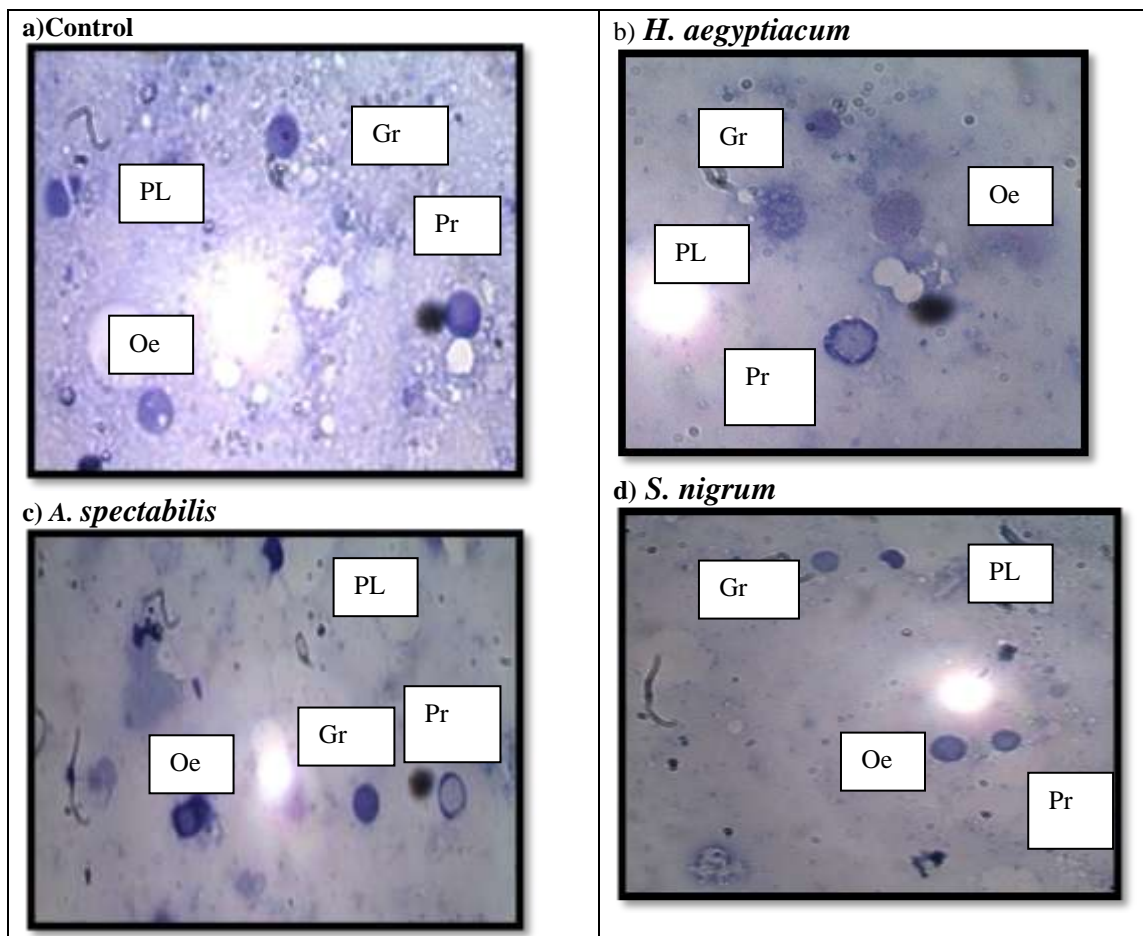


Figure 3. The effect of methanolic plant extracts from *S. nigrum*, *A. spectabilis* and *H. aegyptiacum* on the *C. pipiens* larval haemogram

Similar results were reported by Sharma *et al.* (2008) who noticed that the major effect of *Acorus calamus* oil treatment was observed on plasmatocytes and granular haemocytes of *Spodoptera litura* larvae. Gad and El – DaKheel (2009) reported that *Cinnamomum osmophloeum* and *Matricharia chamomella* oils caused significant decrease in the number of Pr, Gr and Pl and increase the number of Oenocytoide. On contrast, treatment with *C. osmophloeum* and *M. chamomella* oils caused a significant increase in all haemocyte types surface areas. Saxena and Tikku (1990) proved that plumbagin treatment caused damage in the haemocytes of *Dysdercus koenigii* and suppression of filopodial elongations of plasmatocytes and granulocytes. Pendey *et al.* (2007) found that treatment 5th instar larvae of *Danaus chrysippus* caused a marked decrease in the number of haemocytes as well as high variation in relative percentage of different haemocyte types.

The results obtained in this study demonstrate the importance of toxic, growth and development-retarding influence of the extracted plant materials specially *S. nigrum* and *A. spectabilis* on *C. pipiens* mosquitoes. Moreover, application of these materials is not likely to leave harmful residues in the environment since they are naturally materials occurring among the local flora.

REFERENCES

- Abbott, W.S.(1925) A method of computing the effectiveness of an insecticide. *J. Econ. Entomol.*, 18, 265-267.
- Abdelgaleil, S. A.M. (2006). Chemical composition, insecticidal and fungicidal activities of essential oils isolated from *Mentha microphylla* and *Lantana camara* growing in Egypt. *Alex. Sci. Exchange* J.27:18-28.
- Al-Doghairi, M.; A. El-Nadi; E. Elhag and H. Al-Ayedh (2004). Effect of *Solenostemma argel* on oviposition, egg hatchability and viability of *Culex pipiens* L. larvae. *Phytother Res.*, 18 (4): 335-8.
- Arnold, J. W. and Hinks, H. (1983). Haemopoiesis in Lepidoptera. The multiplication of circulation haemocytes. *Can.J.Zoo.* 154:1003-1012.
- Boussaada, O.; M.K. Ben Halima; S. Ammar; D. Haouas; Z. Mighri and A. N. Helal (2008). Insecticidal activity of some Asteraceae plant extracts against *Tribolium confusum*. *Bull. Insectology*, 61(2), 283-289.
- Carlini, CR. and MF. Grossidi-de-Sa (2002). Plant toxic proteins with insecticidal properties. A review on their potentialities as insecticides. *Toxicon*. 40, 1515-1539.
- Cetin, H.; I. Cinbilgel; A. Yanikoglu and M. Gokceoglu (2006). Larvicidal activity of some Labiatae (Lamiaceae) plant extracts from Turkey. *Phytother. Res.*, 20 (12): 1088-90.
- Chitra, K. C.; R. Janarghan; R. Kameswara and K., Nagaiah (1993). Field evaluation of certain plant products in the control of Brinjal Pest Complex. *Indian Journal of Entomology*. 55(3): 237-240.
- Dary, O.; G. P. Georghiou; E. Parsons and N. Pasteur (1990). Microplate adaptation of Gomori's assay for quantitative determination of general esterase activity in single insects. *J. Econ Entomol.*, 83 (6): 2187-92.
- Das, N.G.; I. Baruah; P.K. Talukadar and S.C. Das (2003). Evaluation of botanicals as repellents against mosquitoes. *J. Vect. Borne Dis.*, 40, 49-53.
- Edwards, J. F.; S. Higgs and B. J. Beaty (1998). Mosquito feeding-induced enhancement of Cache Valley Virus (Bunyaviridae) infection in mice. *J. Med. Entomol.*, 35 (3): 261-5.
- Essawy, M. M. (1990). Changes in different haemocyte counts of the last larval instar of *Spodoptera littoralis* (Boisd.) during wound healing. *Alex. Sci. Exch.* 11(4): 151-176.
- Finney, D.J. (1971). Probit analysis 3rd edition. Cambridge University press. Cambridge, 318pp.
- Gad Abir, A , El –DaKheel, Abeer A(2009). Larvicidal activities of *Cinnamomum osmophloeum* and *Matricharia chamomella* extracts against the filarial mosquito *Culex quinquefasciatus* (Diptera:Culicidae)and their effects on its haemogram. *Egypt. Sci. Mag.* 6 (1) PP, 8 - 14.
- Jang, Y. S.; J. H. Jeon and H. S. Lee (2005). Mosquito larvicidal activity of active constituent derived from *Chamaecyparis obtusa* leaves against 3 mosquito species. *J. Am. Mosq. Control Assoc.* , 21(4): 400-3.
- Khater, H. F. and A. A. Shalaby (2008). Potential of biologically active plant oils to control mosquito larvae (*Culex pipiens*, Diptera: Culicidae) from an Egyptian locality. *Rev Inst Med Trop Sao Paulo.*, 50 (2): 107-12.
- Kundu, B. R.; R. Ara; M.M. Begum and Z.I. Sarker (2007). Effect of *Bishkatali Polygonum hydropiper* L. plant extracts against the red flour beetle, *Tribolium castaneum* Herbst. *Univ. J. Zool. Rajshahi Univ.* 26, 93-97.
- Massoud, A. M and I. M. Labib (2000). Larvicidal activity of *Commiphora molmol* against *Culex pipiens* and *Aedes caspius* larvae. *Journal-of-the-Egyptian-Society-of-Parasitology.*, 30 (1): 101-115.
- Michaelakis, A.; A. T. Strongilos; E. A. Bouzas; G. Koliopoulos and E. A. Couladouros (2009). Larvicidal activity of naturally occurring naphthoquinones and derivatives against the West Nile virus vector *Culex pipiens*. *Parasitol Res.*, 104 (3): 657-62.
- Michaelakis, A.; A. P. Mihou; G. Koliopoulos and E.A. Couladouros (2007). Attract-and-kill strategy. Laboratory studies on hatched larvae of *Culex pipiens*. *Pest Manag. Sci.* 63: 954-959.
- Pendey, J P , Upandhyay, AK , Tiwarki, RK (2007). Effect of some plant extracts on haemocyte count and moulting of *Danaus chrysippus* larvae. *J. Adv. Zool.* 28 (1) PP, 14-20.

- Radwan, M. A.; S.R. El-Zemity; S.A. Mohamed and S. M. Sherby (2008). Larvicidal activity of some essential oils, monoterpenoids and their corresponding *N*-methyl carbamate derivatives against *Culex pipiens* (Diptera: Culicidae) Int. J. Trop. Insect Sci. 28:61-68.
- Raymond, M.; C. Berticat; M. Weill; N. Pasteur and C. Chevillon (2001). Insecticide resistance in the mosquito *Culex pipiens*: what have we learned about adaptation? *Genetica.*, 112-113: 287-96.
- Saxena, BP and Tikku, K (1990). Effect of plumbagin on haemocytes of *Dysdercus koenigii* F. J. Animal Sciences. 99(2), PP. 119-124.
- Schmutterer, H.(1995). The neem tree *Azadirachta indica* A. Juss and other meliaceous plants, VCH publisher, Weinheim, Germany, 696.
- Severini, C.; R. Rom; M. Marinucci and M. Rajmond(1993). Mechanisms of insecticide resistance in field populations of *Culex pipiens* from Italy. J. Am. Mosq. Control Assoc., 9, 164-186.
- Sharma, P.R.; Sharma, O.P. and Saxena, B.P. (2008). Effect of sweet flag rhizome oil (*Acorus calamus*) on hemogram and ultrastructure of hemocytes of the tobacco armyworm, *Spodoptera litura* (Lepidoptera: Noctuidae). J. Micron. 39(5), PP. 544-551.
- Smartt, C. T. and J. S. Erickson (2008). CNAc-1 gene is differentially expressed in the subtropical mosquito *Culex nigripalpus* (Diptera: Culicidae), the primary West Nile Virus vector in Florida. *J. Med. Entomol.*, 45 (5): 877-84.
- Srinivasan, R.; J. M. Alvarez; F. Cervantes (2013). The effect of an alternate weed host, hairy nightshade, *Solanum sarrachoides* (Sendtner) on green peach aphid distribution and Potato leafroll virus incidence in potato fields of the Pacific Northwest Crop Protection 46 (2013) 52-56.
- Traboulsi, A. F.; S. El-Haj; M. Tueni; K. Taoubi; N. Abi Nader and A. Mrad(2005). Repellency and toxicity of aromatic plant extracts against the mosquito *Culex pipiens molestus* (Diptera: Culicidae). Pest Manag. Sci. 61: 597-604.
- Venkatachalam, MR. and J. Jenbanesan (2001) Larvicidal activity of *Hydrocotyle javanica* Thumb. (Rutaceae) extract against *Culex quinquefasciatus*. J. Ex. Zool., India, 4 (1), 99-101.
- Xiaorong, T. and H. Taiping (2008). Separation and identification of botanical insecticide 7-hydroxycoumarin and its biological activity against *Aphis craccivora* and *Culex pipiens pallens*. *Nat Prod Res.*, 22 (4): 365-70.
- Zhu, J.; X. Zeng; Yanma; T. Liu; K. Qian; Y. Han; S. Xue; B. Tucker; G. Schultz; J. Coats; W. Rowley and A. Zhang (2006). Adult repellency and larvicidal activity of five plant essential oils against mosquitoes. *J. Am. Mosq. Control Assoc.*, 22 (3): 515-22

الملخص العربي

تأثير بعض المستخلصات النباتية على موت وتطور وخلايا دم يرقات بعوض الكيولكس بيبينز

حسام الدين مجدي زهران، عبير عبد المجيد جاد

وكذلك قللت من تعذير اليرقات وتحولها إلى حشرات كاملة. وقد أظهر مستخلص عنب الديب أكبر تأثير حيث كانت نسب التعذير 18.1% بينما نسبة خروج الحشرات الكاملة كانت 8.5% وذلك عند التركيز 100 جزء في المليون. وقد وجد أيضا أن مستخلصات كلا من الأكوكانثيرا وربيب الشمس المصري كانت فعالة عند التركيزات الأكثر 100 جزء في المليون. وعلاوة على ذلك وجد أن المستخلصات عنب الديب، الأكوكانثيرا وربيب الشمس المصري لها تأثير على أعداد خلايا الدم اليرقات كما أنها أدت إلى انخفاض في مساحة سطح الخلايا بدرجة ملحوظة.

ومن هنا نجد أن المعاملة بهذه المستخلصات في بيئة نمو اليرقات تعطي نتائج أفضل من استخدام المبيدات الكيميائية التي تسبب ضررا للبيئة والإنسان والحيوان وكذلك قد تبدي الحشرات مقاومة لها.

تم دراسة النشاط الإبادي للمستخلص الميثانولي لكل من عنب الديب، الأكوكانثيرا وربيب الشمس المصري وذلك ضد يرقات وتطور بعوضة الكيولكس بيبينز. أظهرت المستخلصات النباتية المختلفة تأثيرات بيولوجية مختلفة وكان الأكثر تأثيرا هو مستخلص نبات عنب الديب والذي أظهر فعل حاد بعد يومين وثمانية أيام من المعاملة حيث كان التركيز المميت 50% من اليرقات 130,8 جزء في المليون على التوالي بعد 48. كانت نسبة الموت في اليرقات 67.5% عند المعاملة بمستخلص عنب الديب و66.5% عند المعاملة بمستخلص الأكوكانثيرا عند التركيز 500 جزء في المليون. إنخفض فقس البيض بصورة غير واضحة في حالة المعاملة بمستخلص عنب الديب. كل التركيزات المستخدمة من مستخلصات كلا من عنب الديب، الأكوكانثيرا وربيب الشمس المصري عائقا واضحا في تطور اليرقات