

Regeneration and Micropropagation Systems of (*Stevia rebaudiana*) var. *sponti*

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ABSTRACT

In order to establish an effective regeneration system of *Stevia (Stevia rebaudiana) var. sponti*, different concentrations of phytohormones were added to MS basal medium individually or in combination with (IAA, BAP and NAA) as callus induction media. The obtaining results indicated that MS basal medium which containing 0.5 mg/l of NAA and 0.5 BAP was the most efficient and effective medium for callus induction.

For shoot regeneration, different recombination of phytohormones were used, addition of NAA (0.1 mg/l) and BAP (1.0 mg/l) to MS basal medium under light condition caused high increase in shooted calli percentage.

Vigor root system was developed after the addition of 3 mg/l of IBA to MS basal medium, while the addition of high concentration of sucrose (80 g / L) found to be ineffective factor for root regeneration and growth. Micropropagation system was performed by exposure of axillaries and terminal buds to solid MS medium which contained 0.1 IAA and 1.0 BA. Then, clusters were cut into small number of plantlets in MS liquid media without any hormone. This method was found to be more effective than using hormonal medium for micropropagation.

INTRODUCTION

Stevia rebaudiana is a herb belong to family Asteraceae family indigenous the higher elevations of Northern Paraguay near the Braziliam borders (Soejarto *et al.*, 1983). This plant has gained importance as a crop for the pharmaceutical and food industries as a result of non-caloric sweeteners extracted from its leaves, mainly stevioside, somatic embryogenesis has been described for more than a hundred plant species (Terzi and Loschiavo, 1990), but the number of reports of somatic embryogenesis among members of the Asteraceae family is still low (May and Trigiano, 1990). The development of protocols for regeneration of *Stevia rebaudiana* via somatic embryogenesis is important as well as the clonal propagation technique of this plant, or as explant material for protoplast isolation and regeneration (Puite, 1992).

The main goal of this investigation is: (1) to

develop a protocol for the induction of somatic embryogenesis from leaf explants of *Stevia rebaudiana*. Also, to establish a most effective regeneration system required for somaclonal variation. (2) To achieve an efficient micropropagation system for the improved plants.

MATERIALS AND METHODS

Plant material

Stevia (Stevia rebaudiana) var. sponti was used as explants donor for this work.

Methods:

1- Tissue culture:

a- Callus induction

Young leaves of *Stevia (Stevia rebaudiana) var. sponti* were cut, then sterilized in sodium hypochlorite solution 2 % for 20 min. and washed twice in sterilized distilled water. Small pieces (about 1 cm²) of leaves were exposed to callus induction media which contained basal MS medium with different hormonal supplements (table 1). Finally, cultures were maintained in incubator in the dark at 25° C until callus production had achieved.

2, 4-D: 2, 4- Dichlorophenoxy acetic acid, NAA: Naphthalene acetic acid, BAP: Benzyl amino purine.

a- Regeneration

- shoot regeneration:

Embryogenic calli were transferred to shoot regeneration medium MS supplemented with 30 g sucrose and phytohormones (0.1, 0.5 and 1.0 mg / L of NAA and 0.1, 0.5 and 1.0 mg / L of BAP).

- Root regeneration:

Two kinds of media were applied, the first composed of MS basal medium supplemented with 30 g sucrose and contained different concentrations of IBA (Indole butaric acid) hormone (1, 2, 3 and 4 mg / L). The second is MS medium (hormone free) that contained high concentration of sucrose (80 g / L)

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Received August 29, 2007, Accepted September 30, 2007

and it was used to induce roots.

Table 1. Different callus induction media.

Supplements added to MS	Concentration (mg/l) and medium designation			
	M1	M2	M3	M4
BAP	0.0	0.1	0.5	0.5
2,4-D	0.0	-	0.5	-
NAA	0.0	0.1	0.5	0.5
Sucrose	30 g/l	30 g/l	30 g/l	30 g/l
Phytigel	6 g	6 g	6 g	6 g
pH	6.8	6.8	6.8	6.8

C- Micropropagation:

Terminal and auxiliary buds were experimented for solid MS medium which contained 0.1 IAA and 1.0 BA in dark for 3 days then transferred to the light (about 1500 Lux) under photosynthesizing 16 h light and 8 h dark at 27 C

2- Plant Multiplication:

Growing clusters (about 3 cm height) were transferred into liquid medium without any addition of hormones (hormone free). The obtaining cultures were cut into 2-3 plant and distributed into new gars contains fresh multiplication medium. This step is replicated every two weeks.

RESULTS AND DISCUSSION

Callus induction:

Embryogenic white calli with compact cells in globular forms and capable of plant regeneration were produced (Fig. 1). Interestingly, hormonal contents in cultural medium were the main factor influenced callus induction and rate of embryogenic type. The present results revealed that the M4 medium (which contained 0.5 mg/l of NAA and 0.5 BAP) was the most efficient for this purpose. The dependence of embryogenic calli on hormonal content was demonstrated in many several works (Filho *et al.*, 2000, Sivaram and Mukunda., 2003).

Maintenance of embryogenic calli:

Subculture of embryogenic calli could be maintained on M4 medium for more than two weeks with the maintaining of their embryogenic ability. Embryogenic calli could be maintained for three months on callus induction medium. The same results were reported by Sanders (1997).

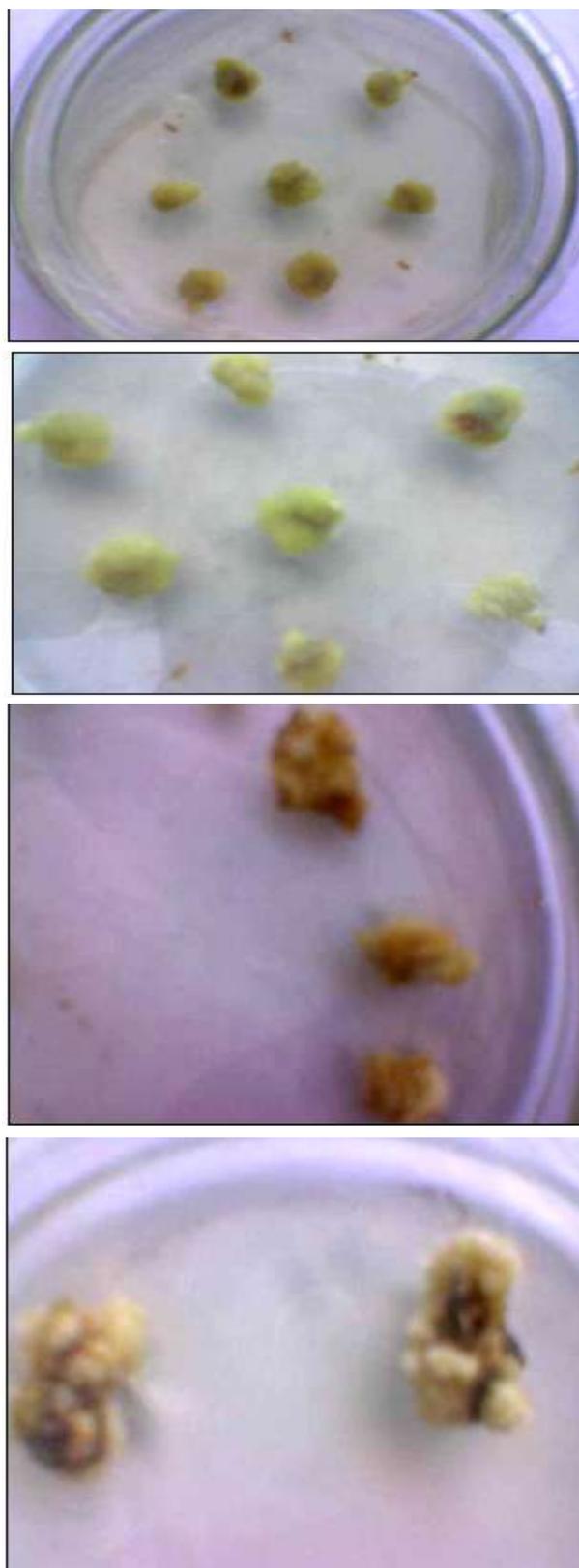


Figure 1: Embryogenic calli proliferation.

Table 2. illustrates the percentage of calli induction in different types of medium under different hormonal concentrations.

Medium Types	M1	M2	M3	M4
Hormonal concentrations	0.0 mg/l 2,4-D, BAP and NAA	0.1 mg/l BAP and 0.1 NAA	0.5 mg/l 2,4-D, BAP and NAA	0.5 mg/l BAP and 0.5 NAA
Calli Percentage	55	64	72	88

Regeneration:

Shoot were producing by addition of NAA (0.1 mg/l) and BAP (1.0 mg/l) to MS basal medium under light condition (Fig. 2). It was noticed that different root system were produced after different concentration of IBA added, increasing in root induction and root growth was detected for addition of 3 mg/l of IBA (Fig. 3). Similar findings was reported by Barathi *et al.*, 2003 and Sivaram and Mukunda (2003) by adding similar hormones to MS medium to produce shoot and root system. Our obtaining results were in agreement of Barathi *et al.*, 2003 established a successful multiplication system for stevia based on culture on free hormone medium. Contrary to results reported by Scoot *et al.*, (2000) high concentration of sucrose had not induced root on the shoot of this variety.



(a)



(b)

Figure 2. Shoot regeneration:

a- shoot formation.

b- shoot growth.



(a)



(b)

Figure 3. Root regeneration:

a- Root formation.

b- Root growth.

Calli percentage

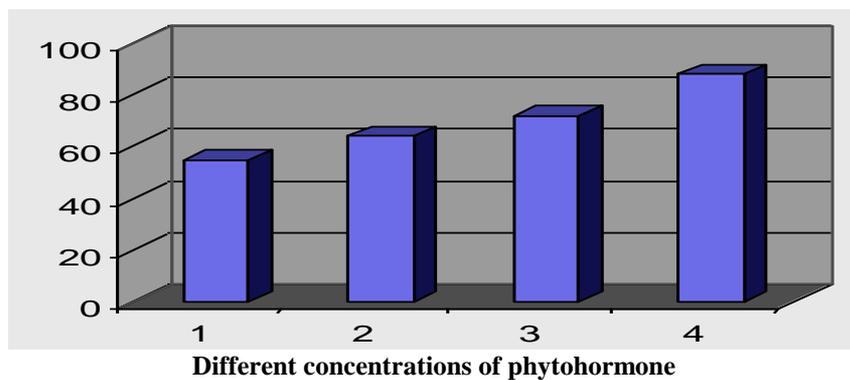


Figure 4. Histogram of calli percentage for different concentrations of phytohormones.

1- 0.0 mg/l 2, 4-D, BAP and NAA

2- 0.1 mg/l 2, 4-D, BAP and NAA

3- 0.5mg/l 2, 4-D, BAP and NAA

4- 1.0 mg/l 2, 4-D, BAP and NAA

Shoot Percentage

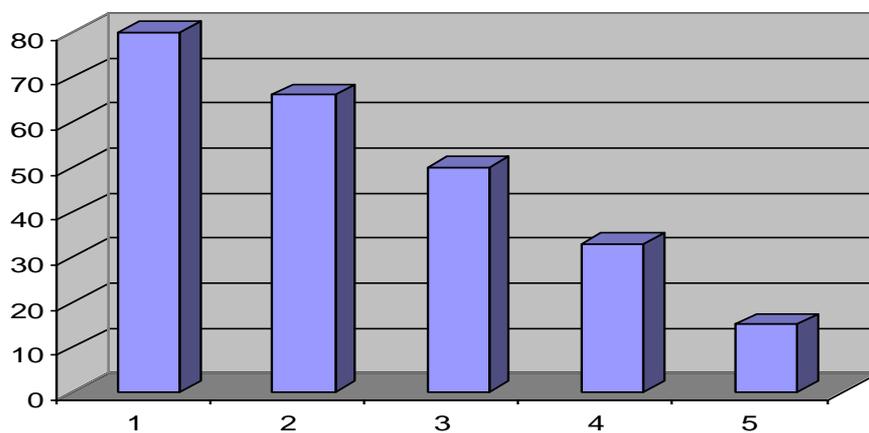


Figure 5. Histogram of shoot percentage for different concentrations of phytohormones.

Where:

1- NAA 0.1 + 1.0 BAP.

2- NAA 0.1 + 0.5 BAP.

3- NAA 0.1 + 0.1 BAP

4- NAA 0.5 + 0.1 BAP

5- NAA 1.0 + 0.1 BAP

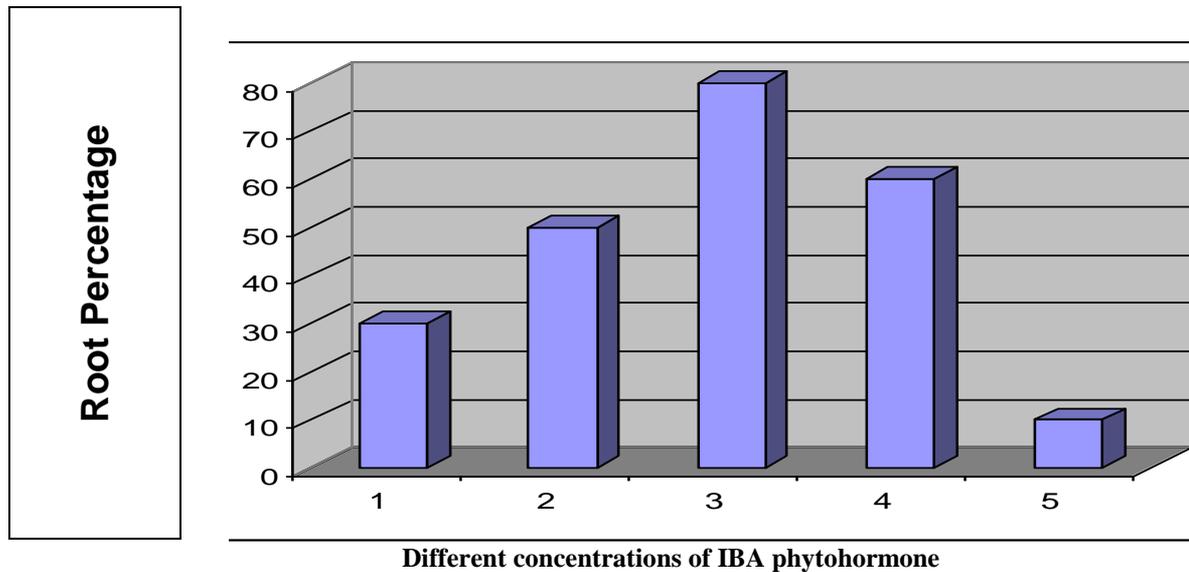


Figure 6. Histogram of root percentage for different concentrations of IBA phytohormones.

1- 1.0 mg/l of IBA + 30 g sucrose

2- 2.0 mg/l of IBA+ 30 g sucrose

3- 3.0 mg/l of IBA+ 30 g sucrose

4- 4.0 mg/l of IBA+ 30 g sucrose

5- 80 g sucrose

REFERENCES

- Barathi V., Peleg K. and Ziv M. (2003). Morphological control and mensuration of potato plantlets from tissue cultures for automated micropropagation. *Plant Cell Tiss. Org. Cult.* 36:331-338.
- Filho J.C, Hashimoto J M and Vieira L .G. (2000). Area de Ecofisiologia, Instituto Agronômico do Paraná. *Canadian Journal of Botany*, 67:970-976.
- May, R.A.and Trigiano, R.N. (1991). Somatic embryogenesis and plant regeneration from leaves of *Dendranthema grandiflora*. *Journal of American Society for Horticultural Science*, 116(2):366-371,.
- Puite, K.J. (1992). Progress in plant protoplast research. *Physiologia Plantarum*, 85:403-410.
- Sanders .C. McWilliams E.L. and Davies F.T. (1997) *In vitro* adventitious callus formation in Asian jasmine (*Trachelospermum asiaticum*). I. Comparative morphology. *J. Amer. Soc. Hort. Sci.* 118:902-905.
- Scoot O., Lewis M., Brackpool A. & Blakesley D. (2003). Auxin and adventitious root initiation in *Eucalyptus globulus*. Abstracts VIIIth International Congress of Plant Tissue and Cell Culture.
- Soejarto, D.D.; Compadre, C.; Medon, P.J. Kamath, S.K. and Kinghorn, A.D.(1983). Potential sweetening agents of plant origin. II. Field research for sweet-tasting *Stevia* species. *Economic Botany*, 37:71-79.
- Sivaram J.C and Mukunda A.A (2003). Tissue culture technique as a tool for *Stevia* improvement. Annual Report Research development Council, 3-9 Taiwan sugar Crop., Taiwan (In Chinese).
- Terzi, M. and Loschiavo, F. (1990). Somatic embryogenesis. In: Bhojwani, S.S. *Plant Tissue Culture, Applications and Limitations; Development in Crop Science*. Elsevier. P.54-66.
- Wada, Y; Tamura, T.; kodama, T.; yamaki, T. and Uchida, Y. (1981). Callus cultures and morphogenesis of *Stevia rebaudiana* Bertoni. *Yukagawa*, 36(4), 215-219.

الملخص العربي

نظم اعادة التشكل و الاكثار السريع لنبات الاستيفيا (صنف sponti)

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و 1.0 مل جرام من بنزاييل امينو بيورين قد تسبب في زيادة نسبة انتاج المجموع الخضرى بصورة معنوية و ايضا فان اضافة 3 مل جرام/ للتر من اندول حمض البيوتاريك تسبب قى زيادة واضحة في انتاج المجموع الجدرى كذلك فان اضافة تركيزات عالية من السكروز (80 جرام) وجد انة عامل غير فعال في أنتاج و نمو الجذور. و للحصول على عدد كبير من النباتات تم استخدام تكتيك التكاثر الدقيق والذي يتم فيه زراعة البراعم الطرفية و الابطية على بيئة MS الصلبة و المحتوية على 1 مل جرام من اندول حمض الخليك و 1.0 مل جرام من حمض البيوتاريك و بعد ذلك تم تقسيم الكتلة الناتجة لعدد من النباتات توضع في بيئة سائلة بدون هرمون.

اجريت هدة الراسة لتحديد الانواع و التراكيب و التركيزات المناسبة من منظمات النمو الاكثر فاعلية في انتاج الكالس و المجموع الخضرى و الجدرى لنبات الاستيفيا (*Stevia rebaudiana*) الصنف sponti .

و قد اوضحت النتائج المتحصل عليها من تجربة عدد من البيئات المختلفة المحتوى الهرمونى ان البيئة الى تتركب من املاح ال MS و المضاف اليها كل من 0.5 مل جرام من اندول حمض الخليك و بيوتاريك امينو بيورين كمنظمات للنمو هى البيئة الاكثر فاعلية و الاعلى في نسبة انتاج الكالس. و باستخدام عدد من البيئات المختلفة في محتواها الهرمونى. تحت ظروف الاضاءة فقد تم التأكد من ان اضافة كل من 1 مل جرام من اندول حمض الخليك